Protease Activity in Plant Tissues (VII)

著者	UCHIKOBA Tetsuya, ODA Mariko, KANEMARU Kaoru, KANEDA Makoto
journal or	鹿児島大学理学部紀要.数学・物理学・化学
publication title	
volume	25
page range	53-58
別言語のタイトル	種々の植物組織のプロテアーゼ活性について (\Ⅱ)
URL	http://hdl.handle.net/10232/00012476

Rep. Fac. Sci. Kagoshima Univ., (Math., Phys. & Chem.), No. 25, 53-58, 1992

Protease Activity in Plant Tissues (VII)

Tetsuya UCHIKOBA¹⁾, Mariko ODA¹⁾, Kaoru KANEMARU¹⁾ and Makoto KANEDA¹⁾

Abstract

Caseinolytic activity of various plant tissues was examined. Favorable protease activity was found in the extracts of rhizome of Baboonroot, *Babiana stricta* Ker-Gawler, seeds of Radish, *Raphaus sativus* L. var. hortensis and rhizome of Ixia, *Ixia hybrida* Hort. Key words: Plant protease, protease.

Introduction

Typical plant proteases so far isolated have belonged mainly to the cysteine protease group. In the plant proteases, enzymatic properties of papain [EC 3. 4. 22. 2] (1) have been considerably investigated. We recently isolated a cysteine protease from sarcocarp of pokeweed, *Phytolacca americana* (2). The enzyme was different from papain in the substrate specificity for some synthetic substrates (2). As the continuation of our previous papers (3-8), we attempted the screening test to find a new type plant protease.

Experimental

Foliage plants were obtained from flower shops, fruits and cereals were purchased from greengrocers and other plants were collected locally in Kagoshima prefecture. Casein was a product of E. Merck, Darmstadt, West Germany; Other reagents were purchased from Wako Pure Chemical Industries Ltd.

Preparation of Sample Solution for Caseinolytic Activity Assay-All samples were ground in equal weight of 0.067 M phosphate buffer, pH 7.3, in a mortar. The homogenates were stirred for 5 min and filtered through a cotton cloth or centrifuged for 10 min at $3,000 \times g$.

The extracts were diluted to the point of appropriate concentration for assay with 0.067 M phosphate buffer, pH 7.3. Characterization of cysteine protease was performed by adding 1 mM L-Cysteine to the sample solution.

Assay of Protease-Proteolytic activity was measured by the method of Kunitz (9) with casein as a substrate. One ml of sample solution was preincubated for 10 min at

¹⁾Department of Chemistry, Faculty of Science, Kagoshima University Kagoshima 890, Japan

Plant	Plant parts	Activity (Units)	
Akagashi, Japanese Evergreen Oak	Nut	0	
(Quercus acuta Thunb.)			
Ashitaba	Leaf, Stem	3	
(Angelica utilis Makino)	,	, i i i i i i i i i i i i i i i i i i i	
Banana, Banana	Fruit	0	
(Musa nana Lour.)			
Banreishi, Sugar apple	Sarcocarp	4	
(Annona squamosa L.)	F	-	
Daikon, Radish	Young Leaf, Stem	9	
(Raphanus sativus L.)	Young root	4	
(Raphanus sativus L. var. hortensis)	Seed	41	
Dorian, Durian	Sarcocarp	11	
(Durio zibethinus DC.)	Sanoodarp		
Esharotto, Shallot	Root	5	
(Allium cepa L vars. aggregatum		Ū	
G. Don, multiplicans L.H. Bailey, solaninum Alef.)			
Gimunema shirubesuta	Leaf	25	
(Gymunema sylvestre, R. Br.)			
Gobou, Great Burdock	Root	0	
(Arctium lappa L.)			
Hakka, Peppermint	Leaf	0	
(Mentha piperata L.)			
Higanbana	Rhizome	0	
(Lycoris radiata Herb.)			
Hiiragi	Berry	0	
(Osmanthus ilicifolius Mouill.)	·		
Hozakiayame, Babiana, Baboon-Root	Rhizome	180	
(Babiana stricta Ker-Gawler)			
Jakkufurutsu, Paramitsu, Jackfruit	Sarcocarp	5	
(Artocarpus heterophyllus Lam.)	Ĩ		
Kaidou, Kaido Crab-Apple	Berry	0	
(Malus micromalus Makino)	·		
Kanariyashi, Canary Island Date Palm	Berry	4	
(Phoenix canariensis hort. ex Chabaud)	·		
Kinmokusei	Sarcocarp	0	
(Osmanthus fragrans var. aurantiacus)			
Kobushi	Sarcocarp	2	
(Magnolia kobus DC.)	r	-	
Komikansou	Fruit	0	
(Phyllanthus urinaria L.)	Leef	0	

Table 1. Caseinolytic Activity of Extracts from Plant Tissues

(Continued on the following page)

Plant	Plant parts	Activit (Units
Kouhii, Coffee	Berry	17
(Coffea arabica 'Blue Mountain')		
Kunugi	Nut	0
(Quercus acutissima Carruth.)		
Kusanoou	Leaf, Stem	0
(Chelidonium majus var. asiaticum)		
Mangosuchin, Mangosteen	Sarcocarp	0
(Garcinia mangostana L.)		
Marubachishanoki	Sarcocarp	10
(Ehretia dicksonii Hance var. japonica Nakai)		
Matsubabotan, Rose Moss	Stem, Leef	2
(Portulaca grandiflora Hook.)	Berry	2
Mube	Fruit	0
(Stauntonia hexaphylla (Thunb.)		
Decne.)		
Nagaimo, Chinese Yam	Bulb	11
(Dioscorea batatas Cecne.)		
Nankinhaze, Chinese tallow tree	Sarcocarp	0
(Sapium sebiferum Roxb.)		
Nasu, Egg plant	Berry	1
(Solanum melongena L.)		
Negi, Ciboule	Leaf	2
(Allium fistulosum L.)		
Nejibana	Leaf, Stem	0
(Spiranthes sinensis var. amoena)	Root	0
Nikuzuku, Common Nutmeg	Seed	0
(Myristica fragrans Houtt.)		
Ninjin, Carrot	Leaf	0
(Daucus carota L. var. sativa DC.)		
Ninniku, Garlic	Rhizome	7
(Allium sativum var. japonicum Kitam.)		
Nira, Chinese chive	Leaf	0
(Allium tuberosum Rottl. ex. K. Spreng.)		
Okahijiki	Leaf, Stem	16
(Salsola komarovii IIjin)		
Orandaayame, Dutch Iris	Rhizome	0
(Iris hollandica hort.)		
Ougonkazura	Leaf	0
(Scindapsus aureus Engl. var. Golden Pothos)		

(Continued on the following page)

(from Table 1.)

Plant	Plant parts	Activit (Units
Outou, Brack cherry	Sarcocarp	5
(Prunus serotina J. F. Ehrh)	Survourp	Ũ
Safuran, Saffron crocus	Rhizome	14
(Crocus sativus L.)		
Sangojyu	Berry	5
(Viburnum odoratissimum Ker-Gawl.)	·	
Sarutoriibara	Sarcocarp	19
(Smilax china L.)		
Satoimo	Bulb	0
(Colocasia antiquorum Schott var. esculenta)		
Satoudaikon, Sugar Beet	Bulb	0
(Beta vulgaris L. var. rapa Dumort.)		
Sendan, Japanese bead-tree	Leaf	10
(Melia azedarach L. var. japonica Makino)		
Serori, Celery	Leaf, Stem	3
(Apium graveolens L.)		
Shishitou, Red pepper	Sarcocarp	11
(Capsicum annuum var. grossum Sendtn. 'Shishitou')		
Suisen, Grand Emperor	Rhizome	14
(Narcissus tazetta L. var. chinensis Roem.)		
Sumomo, Japanese plum	Sarcocarp	0
(Prunus salicina 'Sugar Prume')	-	
Tachinatamame, Jack bean	Sarcocarp	0
(Canavalia glandiata var. ensiformis DC.)		
Tomato, Tomato	Sarcocarp	2
(Lycopersicon esculentum Mill)		
Tsuwabuki	Stem	· 0
(Farfugium japonicum (L. f.) Kitam.)		
Tya, Tea	Sarcocarp	0
(Camellia sinensis (L.) O. Kuntze)		
Tyantin, Chinese toon	Sarcocarp	0
(Cedrela sinensis Juss.)		
Urokomizugoke	Whole	3
(Sphagnum squarrosum Crome)		
Yarizuisen, Ixia	Rhizome	31
(Ixia hybrida hort.)		

	Part	Activity (Units)	
Plant		With Cysteine	Without Cysteine
Kunugi	Nut	64	0
(Quercus acutissima Carruth.)			
Inubiwa	Sarcocarp	25	13
(Ficus erecta Thunb.)			

Table 2. Effect of Cysteine against Caseinolytic Activity of Plants

 30° , and then added to 1 ml of a solution of 1% (w/v) casein containing 0.067 M phosphate buffer, pH 7.3, at 30° . After incubation for 30 min the reaction was terminated by the addition of 3 ml of 5% trichloroacetic acid. After standing for 30 min at room temperature, the precipitate was removed by filtration through Toyo filter paper No. 5C and the absorbancy at 280 nm of the trichloroacetic acid-soluble peptides formed was determined with Hitachi spectrophotometer U-1100.

A unit of activity was defined as that amount which yielded $0.001 A_{280nm}$ unit of change per min in a 1-cm cell under the conditions mentioned above. The specific activity is expressed as the number of enzyme units per 1 ml of juice.

Results and Discussion

The results of the screening test are shown in Table 1.

Caseinolytic activity was observed in several plants. The extracts of rhizome of Baboonroot, *Babiana stricta* Ker-Gawler had high proteolytic activity. Subsequently seeds of Radish, *Raphaus sativus* L. var. hortensis and rhizome of Ixia, *Ixia hybrida* Hort displayed more than 30 units. Small protease activity were present in leaf of *Gymnema sylvestre*, R. Br., sarcocarp of *Smilax china* L. and berry of coffee, *Coffea arabica* 'Blue Mountain'. Comparing the activity with sarcocarp of snake gourds (4, 5), the units on this report were not so large. From the data of Table 2., the protease of nut of *Quercus acutissima* Carruth. and sarcocarp of *Ficus erecta* Thunb. (4) were considered cysteine type enzyme. Because the enzyme activity of these plants increased by adding 1 mM cysteine. In former enzyme was especially more sensitive for cysteine.

References

- 1. Arnon, R. (1970) in *Methods in Enzymology* (Perlmann, G. E. & Lorand, L., eds.) 19, 226-244, Academic Press, New York.
- Kaneda, M., Izumi, S., Fukuda, T., Uchikoba, T. & Tominaga, N. (1988) Phytochemistry, 11, 3661-3662.
- Kaneda, M., Yonezawa, H., & Tominaga, N. (1982) Rep. Fac. Sci., Kagoshima Univ., (Math., Phys., & Chem.) 15, 53-55.
- Kaneda, M., Uchikoba, T., Furugen, K., & Tominaga, N. (1985) *Rep. Fac. Sci., Kagoshima Univ.*, (Math., Phys., & Chem.) 18, 59-63.

- Uchikoba, T., Izumi, S., Fukuda, T., Kaneda, M., & Tominaga, N. (1987) Rep. Fac. Sci., Kagoshima Univ., (Math., Phys., & Chem.) 20, 77-79.
- Uchikoba, T., Sata, I., Akiba, H., Ishihara, S., & Kaneda, M., (1988) Rep. Fac. Sci., Kagoshima Univ., (Math., Phys., & Chem.) 21, 105-110.
- Uchikoba, T., Amakatsu, K., & Kaneda, M. (1990) Rep. Fac. Sci., Kagoshima Univ., (Math., Phys., & Chem.) 23 139-145.
- Uchikoba, T., Taira, M., Tokuda, H., Watariguchi, R. & Kaneda, M. (1991) Rep. Fac. Sci., Kagoshima Univ., (Math., Phys., & Chem.) 24, 55-59.
- 9. Kunitz, M., (1947) J. Gen. Physiol. 30, 291-310.

58