

Malocclusion Impairs Cognitive Behavior via AgRP Signaling in Adolescent Mice

1 **Running title: Malocclusion-induced cognitive decline in young**

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22 **Abstract**

23 **Introduction:** Occlusal disharmony induced by deteriorating oral health conditions, such as tooth loss
24 and decreased masticatory muscle due to sarcopenia, is one of the causes of cognitive impairment.
25 Chewing is an essential oral function for maintaining cognitive function not only in the elderly but also
26 in young people. Malocclusion is an occlusal disharmony that commonly occurs in children. The
27 connection between a decline in cognitive function and malocclusion in children has been shown with
28 chronic mouth breathing, obstructive sleep apnea syndrome, and thumb/digit sucking habits. However,
29 the mechanism of malocclusion-induced cognitive decline is not fully understood. We recently
30 reported an association between feeding-related neuropeptides and cognitive decline in adolescent
31 mice with activity-based anorexia. The aim of the present study was to assess the effects of

32 malocclusion on cognitive behavior and clarify the connection between cognitive decline and
33 hypothalamic feeding-related neuropeptides in adolescent mice with malocclusion.

34 **Methods:** Four-week-old mice were randomly assigned to the sham-operated solid diet-fed
35 (Sham/solid), sham-operated powder diet-fed (Sham/powder), or malocclusion-operated powder diet-
36 fed (Malocclusion/powder) group. We applied composite resin to the mandibular anterior teeth to
37 simulate malocclusion. We evaluated cognitive behavior using a novel object recognition (NOR) test,
38 measured hypothalamic feeding-related neuropeptide mRNA expression levels, and enumerated c-Fos-
39 positive cells in the hypothalamus 1 month after surgery. We also evaluated the effects of central
40 antibody administration on cognitive behavior impairment in the NOR test.

41 **Results:** The NOR indices were lower and the agouti-related peptide (AgRP) mRNA levels and
42 number of c-Fos-positive cells were higher in the malocclusion/powder group than in the other groups.
43 The c-Fos-positive cells were also AgRP-positive. We observed that the central administration of anti-
44 AgRP antibody significantly increased the NOR indices.

45 **Discussion:** The present study suggests that elevated cerebral AgRP signaling contributes to
46 malocclusion-induced cognitive decline in adolescents, and the suppression of AgRP signaling can be
47 a new therapeutic target against cognitive decline in occlusal disharmony.

48 1 Introduction

49 Mastication is a sensory-motor function wherein food is chewed, ground, and swallowed (López-
50 Chaichio et al. 2021). Masticatory dysfunction, such as malocclusion, leads to abnormal sensory input
51 and is associated with cognitive impairment in the elderly (Grayson et al. 2015; Weijenberg et al. 2019).
52 Moreover, the relationship between malocclusion and cognitive impairment has become evident in the
53 young and middle-aged. Chewing gum improves cognitive functions, such as word recall and duration
54 of spatial working memory, in the young and middle-aged (Baker et al. 2004; Stephens and Tunney
55 2004; Wilkinson et al. 2002). Malocclusion is a common dental disease in children (Lombardo et al.
56 2020). Children with obstructive sleep apnea-hypopnea syndrome have a higher incidence rate of
57 malocclusion and lower score of cognitive function than the healthy control (Cai et al. 2013); those
58 with chronic mouth breathing develop malocclusion that changes the shape of their face and mouth,
59 and their cognitive and academic abilities decline as compared to the nasal breathing control group
60 (Grippaudo et al. 2016; Kuroishi et al. 2015). Thumb/digit sucking habits raise a child's risk of
61 increased overjet and open bite followed by malocclusion and speech and language delay (Kumar et
62 al. 2022; Ling et al. 2018). In this way, malocclusion-induced cognitive dysfunction is considered to
63 be a serious health hazard not only to the elderly but also the young.

64 Children and adolescents with malocclusion experience substantial psychological discomfort and a
65 negative impact on their oral health-related quality of life (Bittencourt et al. 2017; Chen et al. 2015).
66 Young subjects with malocclusions present with poor responsiveness to the pain-relieving effects of
67 relaxation (Ruscheweyh et al. 2015). Orthodontic patients who require surgical treatment display high
68 levels of anxiety and depression (Hino et al. 2022). Thus, malocclusion is associated with stress, and
69 chronic stress alters cognitive function (Lupien et al. 2009). Young adults subjected to psychosocial
70 stress for 4 weeks exhibit reduced attentional control (Liston et al. 2009). Cumulative life stress,
71 assessed by the Youth Life Stress Interview, reduces executive functions, which are high-level
72 cognitive abilities, such as spatial working memory and cognitive flexibility, in children (Hanson et al.
73 2012). Repeated restraint stress for 7 days impairs cognitive function in novel object recognition
74 (NOR) tests assigned to animals (Yuen et al. 2012).

75 Recently, an association between eating disorders and occlusal disharmony was revealed (Chiba et al.
76 2022). Anorexia nervosa is a serious eating disorder in adolescent women and causes cognitive decline,
77 such as body image distortion (Dalhoff et al. 2019). Adolescent mice with anorexia nervosa display
78 cognitive decline, and central inhibition of agouti-related peptide (AgRP) and neuropeptide Y (NPY)
79 reverses the cognitive decline (Rokot et al. 2021). Thus, cerebral orexigenic peptides may alter
80 cognitive functions in adolescents. Occlusal disharmony impairs cognitive function by upregulating
81 hippocampal cognitive inhibitors in adult mice (Maeshiba et al. 2022). However, to the best of our
82 knowledge, no prior study has investigated the relationship between orexigenic peptides associated
83 with cognitive function and occlusal disharmony in adolescent mice.

84 The NOR test investigates cognitive paradigms based on working memory, attention, anxiety, and
85 novelty preference in rodents lacking reward or punishment (Antunes and Biala 2012). Rodents
86 approach and explore novel objects frequently when they are simultaneously exposed to familiar and
87 novel objects. The NOR test has been used to assess cognitive function in various animal models of
88 Alzheimer's disease, traumatic brain injury, schizophrenia, Parkinson's disease, autism spectrum
89 disorder, and aging (Grayson et al. 2015).

90 The aim of this study was to evaluate the relationship between hypothalamic orexigenic peptides and
91 cognitive behavior deficiency in young mice with occlusal disharmony using the NOR test This study
92 is expected to reveal the central mechanism underlying malocclusion-induced cognitive decline.

93 **2 Materials and Methods**

94 **2.1 Animals**

95 Male C57BL/6J mice (age, 3 weeks; body weight range, 8–12 g) were purchased from Charles River
96 Laboratories Japan Inc. (Tokyo, Japan). They were individually housed in cages at 24 ± 2 °C and $50 \pm$
97 10% humidity under a 12 h/12 h light/dark cycle. The light period was between 07:00 and 19:00. The
98 mice had *ad libitum* access to a sterile standard diet (3.4 kcal/g; CE-2; CLEA Japan Inc. Tokyo, Japan)
99 and water in a pathogen-free facility. All experimental protocols were approved by the Kagoshima
100 University Committee (No. D21035). The present study conformed to Animal Research: Reporting *In*
101 *Vivo* Experiments (ARRIVE) Guidelines v.2.0 for Preclinical Animal Studies. Sample size and
102 inclusion and exclusion criteria are described in the Supplemental Methods.

103 **2.2 Mouse Occlusal Disharmony Model Induction**

104 Mice were housed individually, and acclimated to a rearing environment for 1 week prior to the
105 experiments and randomly assigned the Sham/solid, Sham/powder, or Malocclusion/powder group.
106 Since mice with occlusal disharmony are unable to eat hard foods due to poor bite, a Malocclusion/solid
107 group was not included. Random numbers were generated by Microsoft Excel. The mice in the powder
108 diet-fed groups were acclimated to the powder diet for 3 days before surgery. The occlusal disharmony
109 model of the anterior teeth simulates a stressful situation similar to that seen in humans and induces
110 cognitive impairment (Yoshihara et al. 2009; Yoshihara et al. 2011; Shimizu et al. 2018; Suita et al.
111 2020). Occlusal disharmony was induced according to previously reported methods, with certain
112 modifications (Shimizu et al. 2018; Suita et al. 2020). In brief, the mice were anesthetized by the
113 intraperitoneal (ip) administration of a mixed anesthetic agent: 0.3 mg/kg medetomidine (Meiji Seika
114 Pharma, Japan), 4.0 mg/kg midazolam (Sandoz, Tokyo, Japan), and 5.0 mg/kg butorphanol (Meiji
115 Seika Pharma, Tokyo, Japan). Then, 1.0 mm of composite resin (BEAUTIFIL Flow Plus, SHOFU Inc.,
116 Kyoto, Japan) was applied to their mandibular anterior teeth with pre-treatment of FL-Bond II Primer
117 and Bonding Agent (SHOFU Inc., Kyoto, Japan). Next, light curing was performed for 30 sec at 1200

118 mW/cm² in the vertical direction using a PEN Bright (SHOFU, Kyoto, Japan). The mice were
119 recovered from anesthesia by the ip administration of 0.3 mg/kg atipamezole (Nippon Zenyaku Kogyo,
120 Koriyama, Japan). The sham mice were anesthetized, underwent no intervention, and were recovered.
121 All mice were individually housed for 1 month. Photographs and a schematic representation of the
122 mouse occlusal disharmony model are shown in Figure 1A.

123 **2.3 Food intake and Body Weight**

124 Food intake and body weight of mice in three groups measured daily at 07:00. Body weight gain was
125 calculated during each period. Food intake per body weight was calculated for each experimental day,
126 and body weight gain per food intake between Days 1 and 4, Days 4 and 9, and Days 9 and 30.

127 **2.4 Refeeding Test**

128 The mice used in the refeeding tests differed from those used in the behavior tests. The refeeding tests
129 were performed according to the schedule diagram in Figure 1B. The mice in each group were fasted
130 for 16 h and had *ad libitum* water access. Cumulative food intake was measured 0.5, 1, 2, and 4 h post-
131 feeding.

132 **2.5 NOR Test**

133 The NOR tests were performed according to a previous study (Rokot et al. 2021) and are shown in the
134 schedule diagram in Figure 1B. Each mouse was placed in an empty 60 × 60 × 70 cm box with black
135 walls and an open top for video recording. The mice were allowed to acclimate to the environment for
136 10 min (habituation phase) and returned to their home cages. Two objects of the same color, shape,
137 and size were placed on opposite sides of each box. The mice were placed in the boxes, where they
138 could freely explore for 10 min (Phase I), and returned to their home cages. The objects were removed
139 from the boxes. The mice were placed in the cleaned, empty boxes for 10 min (resting phase) and
140 returned to their home cages. The objects used in Phase I were replaced in the boxes. However, one
141 item was placed in the same position as before (familiar), whereas the other was placed in a different
142 position. The mice were placed in the boxes, allowed to explore freely for 10 min (Phase II), and
143 returned to their home cages. The objects were removed from the boxes. The mice were then placed
144 again in the cleaned, empty boxes for 10 min (resting phase). A familiar object and novel object of a
145 different color, shape, and size were placed in the same positions as in Phase I. The mice were placed
146 in the boxes and allowed to explore for 10 min (Phase III). All objects and the box were cleaned with
147 70% ethanol to remove any residual odors after each phase. Object exploration was defined as touching
148 it with the nose but climbing onto it or chewing it was not considered exploration. In contrast, a mouse
149 that sniffed the object and climbed was considered to have explored it. The NOR index was calculated
150 as follows: (time of exploration of the new object - time of exploration of the familiar object) / (time
151 of exploration of the new object + time of exploration of the familiar object). A schematic diagram of
152 the NOR test procedure is shown in Figure 2A.

153 **2.6 Tissue Sampling**

154 The mice were fasted for 4 h to reduce variability in the expression of their feeding-associated peptides.
155 Tissues and peripheral blood were sampled, as described below, from the various mice subjected to the
156 behavior test. The mice were anesthetized by the ip administration of a mixture of 0.3 mg/kg
157 medetomidine, 4.0 mg/kg midazolam, and 5.0 mg/kg butorphanol. Peripheral blood was collected from
158 the heart. Plasma was separated by centrifugation at 4°C and stored at -80°C until assay. The levels of
159 blood glucose (BG), total cholesterol (T-CHO), and triglyceride (TG) in the plasma samples were

160 measured in Kyudo Co., Ltd. (Saga, Japan). Mice were perfused with 0.1 M phosphate buffer and
161 euthanized by perfusion without awakening from anesthesia. Brain tissues were excised and isolated
162 for RT-qPCR. For immunohistochemistry analysis, the mice were perfused with 0.1 M phosphate
163 buffer (pH 7.0) followed by 4% paraformaldehyde and 0.5% glutaraldehyde in 0.1 M phosphate buffer.

164 **2.7 RT-qPCR**

165 The mice were perfused with 0.1 M phosphate buffer and their hypothalami were isolated. Total RNA
166 was extracted using the RNeasy Plus Mini Kit (No. 74134; QIAGEN, Hilden, Germany). The RNA
167 was reverse transcribed to cDNA using SuperScript IV VILO (No. 11756050; Invitrogen, Carlsbad,
168 CA, USA). RT-qPCR was performed using SYBR Green Master Mix (Thermo Fisher Scientific,
169 Waltham, MA, USA) according to the manufacturer's protocol. The primers used in RT-qPCR are
170 listed in Supplementary Table S1.

171 **2.8 Immunohistochemistry**

172 Coronal sections (25 μ m) of the hypothalami were cut on a cryostat (CryoStar NX70; Thermo Fisher
173 Scientific, Waltham, MA, USA) Hypothalamus sections were incubated with rabbit anti-c-Fos
174 antibody (1:100; ABE457; Merck Millipore, Belize, MA, USA) or mouse anti-c-Fos (1:500; sc-
175 166940; Santa Cruz Biotechnology, Dallas, TX, USA), rabbit AgRP (1:1,000; H-003-53; Phoenix
176 Pharmaceuticals, Burlingame, CA, USA), and guinea pig anti-product gene protein 9.5 (PGP9.5;
177 1:5,000; GP14104; Neuromics, Edina, MN, USA), which is a pan-neuronal marker (Schofield et al.
178 1995, Day and Thompson, 2010), at 4 °C overnight. The sections were then incubated with the
179 secondary antibodies Alexa Fluor 488-conjugated donkey anti-rabbit IgG (1:500; ab150065; Abcam,
180 Cambridge, UK), Alexa Fluor 555-conjugated donkey anti-mouse IgG (1:500; ab150110, Abcam), and
181 Alexa Fluor 647-conjugated donkey anti-guinea pig IgG (1:500; 706-605-148; Jackson
182 ImmunoResearch Labs, West Grove, PA, USA) at 25 °C for 3 h. The nuclei were stained with 4',6-
183 diamidino-2-phenylindole dihydrochloride solution (DAPI; No. D523; Dojindo Molecular
184 Technologies Inc., Kumamoto, Japan). The image was observed using confocal laser microscopy (LSM
185 900; Carl Zeiss AG, Jena, Germany). The c-Fos-positive cells were enumerated on one side of each of
186 six hypothalamus tissue sections per mouse at 200 \times magnification using confocal laser microscopy.
187 The averages were calculated and used in the subsequent analysis. The numbers of c-Fos- and DAPI-
188 positive cells were manually counted, and c-Fos-positive cell numbers were normalized by the
189 respective DAPI-positive cell numbers. AgRP and c-Fos-, DAPI and c-Fos-, AgRP and DAPI-, AgRP
190 and PGP9.5-, and DAPI and PDP9.5-positive cells in arcuate nuclei were observed at 400 \times
191 magnification using confocal laser microscopy.

192 **2.9 Cannula Implantation**

193 The mice were anesthetized by the ip administration of a mixture of 0.3 mg/kg medetomidine, 4.0
194 mg/kg midazolam, and 5.0 mg/kg butorphanol. A guide cannula (25-gauge; Eicom, Kyoto, Japan) was
195 implanted into the right lateral ventricle with a Kopf stereotaxic frame (David Kopf Instruments,
196 Tujunga, CA, USA). The stereotaxic coordinates were 0.8 mm posterior to the bregma, 1.5 mm left
197 lateral to the midline, and 1.2 mm below the outer surface of the skull. The guide cannula was secured
198 with dental cement (Super Bond; Sun Medical Co. Ltd., Moriyama, Japan) and anchored with two
199 stainless steel screws (AN-3; Eicom) fixed to the dorsal surface of the skull. A dummy cannula (AD-
200 4; Eicom) was placed into each guide cannula and fixed with a screw cap (AC-4; Eicom) to prevent
201 occlusion. After the cannula implantation, the mice were recovered from anesthesia by the ip
202 administration of 0.3 mg/kg atipamezole. The mice were lightly anesthetized by isoflurane inhalation
203 and intracerebroventricular (icv) administration was performed. The dummy cannulae were replaced

204 with microinjection cannulae (AMI-5; Eicom) that were 1 mm longer than the guide cannulae. Each
205 microinjection cannula was connected to a polyethylene tube (PE-50; Clay Adams, Parsippany, NJ,
206 USA). At the end of the experiments, the mice were euthanized by carbon dioxide inhalation and the
207 correct locations of the icv cannulae were verified with 10 μ L of 0.05% cresyl violet dye.

208 **2.10 Drug Administration**

209 Anti-AgRP antibody (AF634; R&D Systems, Minneapolis, MN, USA) was dissolved in saline solution
210 (0.9% NaCl) and administered intracerebroventricularly at a dose of 0.1 μ g/mouse from Day 25 to Day
211 30 at 07:00–08:00. Saline solution (2 μ L) was also intracerebroventricularly administered as a vehicle.
212 Specificity of this antibody was verified in previous studies (Cortes-Campos et al. 2013; Okamoto et
213 al. 2016; Fukuhara et al. 2019; Ou et al. 2019; Kim et al. 2020).

214 **2.11 Data Analysis**

215 Data are represented as means \pm standard error of the mean (SEM). Pairwise comparisons between
216 groups were conducted using Student's *t*-test. One-way ANOVA followed by Tukey's multiple
217 comparisons tests was used to compare the three groups. Two-way ANOVA followed by Tukey's or
218 Bonferroni's multiple comparisons tests were used to compare two or three groups over time. Pearson
219 correlation coefficient was used to explore the correlations between variables the variables. Differences
220 were considered statistically significant at $p < 0.05$. All statistical analyses were performed using
221 GraphPad Prism 9 (GraphPad Software, La Jolla, CA, USA).

222 **3 Results**

223 **3.1 Mice with Malocclusion had Lower Body Weight but no Alteration in Food Intake**

224 The mean body weight of the Malocclusion/powder group was significantly lower than that of the
225 Sham/solid group between Days 1 and 30 and Sham/powder group between Days 1 and 12 ($F_{2, 25} =$
226 $9.014, p = 0.0011$, two-way ANOVA; Fig. 1C). The mean body weight of the Sham/powder group was
227 significantly lower than that of the Sham/solid group on Day 26 (Fig. 1C). Food intake was
228 significantly lower in the Malocclusion/powder group than in the Sham/solid and Sham/powder groups
229 on Days 1 and 2 ($F_{2, 25} = 6.166, p = 0.0065$, two-way ANOVA; Fig. 1D). Since the mean body weight
230 of the Malocclusion/powder group was significantly lower than that of the Sham/solid group, food
231 intake/body weight and body weight gain/food intake was calculated to further assess the effects of
232 malocclusion. The mean of food intake/body weight in the Malocclusion/powder group was lower
233 between Days 1 and 2 but higher between Days 4 and 7 than that in the other groups and higher than
234 that in the Sham/powder group on Day 8 (Fig. 1E). However, there were no significant differences
235 observed among the groups between Days 9 and 30 ($F_{2, 25} = 1.382, p = 0.2697$, two-way ANOVA; Fig.
236 1E). Moreover, the mean of body weight gain/food intake in the Malocclusion/powder group was
237 significantly lower than that of the Sham/solid group on Days 1, 21, and 26 as well as that of the
238 Sham/powder group on Days 1, 2, and 21 ($F_{2, 25} = 16.02, p < 0.001$, two-way ANOVA; Fig. 1E).
239 Furthermore, the body weight gain/food intake was significantly lower in the Malocclusion/powder
240 group than that in the other groups from Day 1 to Day 4 ($F_{2, 25} = 15.47, p < 0.0001$, one-way ANOVA;
241 Fig. 1F). Finally, the BG and TG concentrations in peripheral blood showed no differences among the
242 groups (BG: $F_{2, 23} = 1.165, p = 0.3296$, TG: $F_{2, 23} = 0.6130, p = 0.5503$, one-way ANOVA; Fig. 1G).
243 Notably, the T-CHO concentration in the peripheral blood of the Sham/solid group was significantly
244 higher than that of the other groups ($F_{2, 23} = 9.645, p = 0.0009$, one-way ANOVA; Fig. 2G).

245 **3.2 Malocclusion Increased Cumulative Post-Fasting Food Intake in Mice**

246 In the refeeding test, the cumulative food intake was significantly higher in the Malocclusion/powder
247 group than in the Sham/solid and Sham/powder groups after 1 and 2 h ($F_{2, 25} = 3.692$, $p = 0.00394$,
248 two-way ANOVA; Fig. 1F). Nevertheless, there were no significant differences in the cumulative food
249 intake among groups after 4 h (Fig. 1F).

250 **3.3 Malocclusion Impaired Cognitive Function in NOR Test**

251 There were no significant differences in Phase I NOR indices among groups ($F_{2, 25} = 1.420$, $p = 0.26$,
252 one-way ANOVA; Fig. 2B). However, the Phase II NOR index was lower in the Malocclusion/powder
253 group than in the Sham/solid and Sham/powder groups, and the Phase II NOR index was lower in the
254 Sham/powder than in the Sham/solid group ($F_{2, 25} = 39.53$, $p < 0.001$, one-way ANOVA; Fig. 2C). The
255 Phase III NOR index was significantly lower in the Malocclusion/powder group than in the Sham/solid
256 or Sham/powder groups, and the Phase III NOR index of the Sham/powder group reverted to the level
257 of that of the Sham/solid group ($F_{2, 25} = 23.41$, $p < 0.001$, two-way ANOVA; Fig. 2D).

258 **3.4 Hypothalamic mRNA Levels of AgRP and Ucn2 were Increased in Mice with** 259 **Malocclusion**

260 The AgRP and urocortin2 (Ucn2) mRNA levels were significantly higher in the Malocclusion/powder
261 group than in the Sham/solid and Sham/powder groups (AgRP: $F_{2, 25} = 9.574$, $p < 0.001$, Ucn2: $F_{2, 23} =$
262 4.553 , $p = 0.0216$, one-way ANOVA; Fig. 3A). In contrast, the mRNA levels of NPY, POMC
263 (proopiomelanocortin), CART (cocaine- and amphetamine-regulated transcript), CRF (corticotropin-
264 releasing factor), Ucn1, Ucn3, AVP (arginine vasopressin), OXT (oxytocin), and orexin did not
265 significantly differ among groups (Fig. 3A). Interestingly, although the Phase III NOR indices were
266 negatively correlated with mRNA expression levels of AgRP ($r = -0.8166$, $p < 0.01$, Fig. 3B), there
267 was no correlation between Phase III NOR indices and mRNA expression levels of Ucn2 (Fig. 3B).

268 **3.5 Number of c-Fos-Positive Cells in the Arcuate Nucleus of the Hypothalamus was** 269 **Increased in Mice with Malocclusion, and These Cells were AgRP-Positive**

270 We examined the arcuate nuclei of the hypothalami as the AgRP neurons are localized there. There
271 were significantly more c-Fos-positive cells in the arcuate nuclei of the Malocclusion/powder group
272 than in those of the Sham/solid and Sham/powder groups ($F_{2, 18} = 16.68$, $p < 0.001$, one-way ANOVA;
273 Fig. 4A and B). Moreover, the c-Fos-positive cells were AgRP-positive (Fig. 4C).

274 **3.6 Icv Anti-AgRP Antibody Administration Reversed Malocclusion-Induced Cognitive** 275 **Impairment**

276 A previous study demonstrated that Ucn2 has no effect on NOR other than influencing the number of
277 attempts to climb the new object (Clark et al. 2007), and the present study revealed that mRNA
278 expression of AgRP, not Ucn2, had a negative correlation with NOR indices in Phase III (Fig. 3B).
279 For these reasons, we focused on AgRP alone. The Phase I NOR indices did not differ between mice
280 administered antibody and those administered vehicle ($t(8) = 0.4876$, $p = 0.6389$, t -test; Fig. 5A). The
281 Phase II and Phase III NOR indices were significantly reversed in the Malocclusion/powder group
282 administered anti-AgRP antibody by icv for 5 days ($t(8) = 6.534$, $p = 0.0002$; Fig. 5B and $t(8) = 9.629$,
283 $p < 0.0001$, t -test; Fig. 5C). Icv anti-AgRP antibody administration did not alter body weight or food
284 intake relative to vehicle administration ($F_{1, 8} = 1.394$, $p = 0.27$ and $F_{1, 8} = 3.493$, $p = 0.10$, two-way
285 ANOVA; Supplementary Fig. S1).

286 **4 Discussion**

287 The mean body weight of the Malocclusion/powder group was significantly lower than that of the
288 Sham/solid group for 30 days and significantly lower than that of the Sham/powder group for the first
289 11 days. Food intake was significantly lower in the Malocclusion/powder group than in the other
290 groups for the first 2 days. While the food intake/body weight of the Malocclusion/powder group was
291 lower for the first 2 days after the induction of occlusal disharmony, it became higher than that of other
292 groups between Days 4 and 8, returning to normal levels from then on. Moreover, the body weight
293 gain/food intake between Days 1 and 4 was lower in the Malocclusion/powder group than in the other
294 groups. However, this value returned to normal levels between Days 4 and 30. The total cholesterol
295 concentrations in the peripheral blood samples from both powder diet-fed groups were lower than those
296 from the Sham/solid group. However, a decline in the Phase III NOR indices was observed in the
297 Malocclusion/powder group, yet not in the Sham/powder group. Additionally, the decrease in
298 cholesterol showed no effect on body weight gain. A previous study demonstrated that the high
299 cholesterol level seen in an obesity model induced by the Cafeteria diet impairs cognitive function in
300 the NOR task (Lewis et al. 2019). Hence, the resin construction surgery itself had virtually no influence
301 as of the day upon which the behavior tests and tissue samplings were conducted. BG levels in all
302 groups were slightly increased by the anesthesia (Ochiai et al.2016).Occlusal disharmony induced by
303 the composite resin applied on the mandibular incisors reduces body weight (Suita et al. 2020). The
304 present study showed lower body weight in the Malocclusion/powder group than in the Sham/solid
305 group. This finding was consistent with previous studies.

306 In present study both Sham/powder presented with decreased the NOR index in Phase II of NOR test.
307 On the other hand, Malocclusion/powder group presented with decreased both NOR indices in Phase
308 II and III. Occlusal disharmony may lead to abnormal sensory input and is associated with working
309 memory function impairment (Sakatani et al. 2013). The NOR task is considered to evaluate the
310 involvement of working memory in object location and features, memory consolidation, and
311 reorganization of consolidated memory associated with input of new information (Antunes and Biala
312 2012). One of key areas of the brain involved in this processing is the hippocampus (Furini et al. 2020).
313 In a previous study, occlusal disharmony impaired cognitive ability of young mice when performing
314 the NOR task 1 and 4 weeks after the loading (Maeshiba et al. 2022). Moreover, the protein levels of
315 various cognitive suppressor molecules in the hippocampus, such as amyloid- β and phosphorylated tau,
316 were increased at 1 week and were reduced at 4 weeks after the loading (Maeshiba et al. 2022).
317 Anorexigenic peptide signals in the hypothalamus have been reportedly associated with anorexia-
318 induced recognition decline in juvenile mice (Rokot et al. 2021). Therefore, the hypothalamus may
319 also be involved in malocclusion-induced cognitive decline in young mice.

320 Previous study reported that mice fed a powdered diet have lower position recognition test scores than
321 those fed a solid diet and decrease in hippocampal nerve growth-promoting factor BDNF levels
322 (Fukushima-Nakayama et al. 2017). Although Malocclusion/powder group presented the positional
323 recognition decline in the present study, the decline was reversed by central administration of ant-
324 AgRP antibody. The decline in positional recognition induced by malocclusion may have a different
325 mechanism from that induced by a powder diet.

326 Hypothalamic AgRP mRNA was significantly upregulated in the Malocclusion/powder group. AgRP
327 neuron activation occurred in response to malocclusion and was suppressed by central AgRP antibody
328 administration. Upregulated hypothalamic AgRP is associated with significantly lower NOR test scores
329 in mouse anorexia models (Rokot et al. 2021). Likewise, while the AgRP signal produces an aversive
330 condition, its inhibition enhances the learning of a sensory cue-initiated food-acquisition task (Berrios
331 et al. 2019). By blocking AgRP activity via icv antibody administration, the recognition ability of the
332 Malocclusion/powder group may increase compared to that of Sham/solid mice. Chronic,

333 unpredictable, mild stress impaired the recognition ability of rats in the NOR task, and the melanocortin
334 4 receptor, a receptor to which AgRP binds, was upregulated in the nucleus accumbens (Goudarzi et
335 al. 2020). AgRP neurons project to the ventral striatum, including the nucleus accumbens, and
336 contribute to motivation induced by dopamine signals (Reichenbach et al. 2022). The nucleus
337 accumbens is an important area not only associated with the reward system, but also recognition
338 memory such as taste neophobia (Alejandro et al. 2020). Thus, the AgRP signal may be a key mediator
339 of malocclusion-induced cognitive decline.

340 AgRP is a potent cerebral orexigenic peptide (Sohn et al. 2015). The Malocclusion/powder group
341 exhibited no alteration in daily food intake between Day 4 and Day 30. Nevertheless, the cumulative
342 food intake after overnight fasting increased until 2 h and normalized after 4 h compared with the other
343 groups. AgRP and NPY contribute to food intake during 3 h of refeeding after 8 h of fasting (Palou et
344 al. 2009). The observed increase in food intake in the Malocclusion/powder group at 2 h may reflect
345 significant AgRP neuron activation.

346 A prior survey disclosed a negative correlation between malocclusion severity (assessed by the Index
347 of Orthodontic Treatment Need-Dental Health Component) and mastication (assessed by food intake
348 ability) (Choi et al. 2016). The association between eating disorders and occlusal disharmony has been
349 shown (Chiba et al. 2022), and individuals with anorexia display perturbation of cognitive function
350 (Kaye et al. 2008) and elevated plasma AgRP levels (Moriya et al. 2006). Patients with anorexia do
351 not eat even when AgRP is activated (Escelsior et al. 2022). Suppression of AgRP signaling reverses
352 the decline in cognitive function in a mouse anorexia model (Rokot et al. 2021). In the present study,
353 the Malocclusion/powder group exhibited no alteration in daily food intake even though their AgRP
354 neurons were activated. Thus, there may be certain neurophysiological similarities between
355 malocclusion and anorexia. AgRP signaling can be a new therapeutic target for cognitive decline in
356 occlusal disharmony and anorexia nervosa.

357 The present study also showed that Ucn2 mRNA expression was upregulated in the
358 Malocclusion/powder group. Ucn2 is a CRF family peptide that suppresses food intake and gastric
359 motility and is anxiolytic (Martínez et al. 2004). Central Ucn2 infusion alters the frequency with which
360 animals climb onto new objects but has no apparent effect on latency to touch the new object or the
361 total number of touches and climbs onto the new object (Clark et al. 2007). Hence, Ucn2 may have
362 little effect on cognitive function. For this reason, we focused on assessing the effects of AgRP in the
363 present study. The anorexigenic effect of Ucn2 may have contributed to the observed lack of change
364 in food intake in the Malocclusion/powder group despite the increase in levels of the orexigenic peptide
365 AgRP. Although AgRP neurons in the hypothalamus contribute to CRF activity in a fasted state
366 (Fernandes et al.2022), the interaction between AgRP and Ucn2 has not yet been reported. The
367 cognitive impairment in NOR test correlated with mRNA expression of AgRP, not Ucn2, in the present
368 study. In summary, Ucn2 may act independently of AgRP in mice with malocclusion.

369 CRF is a peptide that regulates various stress responses that affect gastrointestinal function and induce
370 anxiety and the secretion of stress-related hormones, such as corticosterone in rodents and cortisol, in
371 humans (Deussing and Chen 2018). Chronic stress may induce other factors besides, or in addition to,
372 CRF (Ataka et al. 2012). Chronic homotypic (but not heterotypic) stress is not associated with any
373 alteration in hypothalamic CRF (Zheng et al. 2010). Although malocclusion is a stressor, no
374 hypothalamic CRF mRNA upregulation was observed in the Malocclusion/powder group here.
375 Malocclusion may be a form of chronic homotypic stress. Plasma corticosterone levels do not change
376 for 4 weeks in a rat malocclusion model (Irie et al. 2011). The duration of malocclusion was 4 weeks

377 in the present study. CRF may not have been implicated in the mechanism of malocclusion-induced
378 cognitive impairment.

379 The present work had certain limitations as we did not perform gene silencing to remove the target
380 mRNA nor did we use knockout mice. Additionally, we did not identify the projection target of AgRP.
381 Further research is needed to more thoroughly explore the complex interaction malocclusion-induced
382 cognitive decline and AgRP signaling. Additionally, the results of this study should be corroborated or
383 validated in future clinical studies. However, the present study is the first to show that AgRP signaling
384 in the arcuate nucleus of the hypothalamus contributes to occlusal disharmony-induced recognition
385 decline. AgRP signaling might be a novel target for the treatment of a defect of cognitive ability
386 induced by the occlusal disharmony.

387 **5 Conflict of Interest**

388 The authors declare that the research was conducted in the absence of any commercial or financial
389 relationships that could be construed as a potential conflict of interest.

390 **6 Author Contributions**

391 J.K. contributed to study conception and design and data acquisition, interpretation, and analysis and
392 drafted and critically revised the manuscript; K.A. and H.I. contributed to study design and data
393 acquisition, interpretation, and analysis and drafted and critically revised the manuscript; Y.O.
394 contributed to study conception and design and data interpretation and drafted and critically revised
395 the manuscript; K.Y., K.M., and T.I. contributed to data interpretation and drafted and critically revised
396 the manuscript; A.A. contributed to study design and data interpretation and critically revised the
397 manuscript; S.M. contributed to study conception and design and data interpretation and critically
398 revised the manuscript. All authors approved the final version of the manuscript and agreed to be
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575 10 Data Availability Statement

576 The original contributions presented in the study are included in the article/supplementary material,
577 further inquiries can be directed to the corresponding author.

578 11 Figure Legends

579 **Figure 1. Photographs and schematic diagram of occlusal disharmony in mice, schedule diagram,**
580 **and body weight and food intake time courses.** (A) Representative image of malocclusion (1-mm
581 increase in vertical height) induced by cementing composite resin onto mandibular incisors of mice.
582 (B) Mice were assigned to Sham operation with solid chow (Sham/solid, $n = 9$), Sham operation with
583 powder chow (Sham/powder, $n = 10$), or Application of composite resin on mandibular anterior teeth
584 with powder chow (Malocclusion/powder, $n = 9$) group. Experiments were performed according to the
585 schedule shown. (C, D) Body weight (C) and food intake (D) were measured for 30 d under
586 experimental conditions. (E, F) Food intake/body weight (E) and body weight gain/food intake from
587 Day 1 to 4, Day 4 to 9, and Day 9 to 30 (F) were calculated. (G) Blood glucose (BG), total cholesterol
588 (T-CHO), and triglyceride (TG) concentrations in peripheral blood were measured. (H) Cumulative
589 food intake was measured in other mice: Sham/solid ($n = 10$), Sham/powder ($n = 10$) and
590 Malocclusion/powder group ($n = 8$) after 16 h fasting. Data are represented as means \pm SEM. In C, D
591 and E, differences were considered significant at $^*p < 0.05$ and $^{**}p < 0.01$ compared with Sham/solid
592 group, and at $^{\#}p < 0.05$ and $^{\#\#}p < 0.01$ compared with Sham/powder group. In F, G and H differences
593 were considered significant at $^*p < 0.05$ and $^{**}p < 0.01$.

594 **Figure 2. Schematic illustration of novel object recognition (NOR) test and NOR indices.** (A) All
595 mice in Sham/solid ($n = 9$), Sham/powder ($n = 10$), and Malocclusion/powder group ($n = 9$) were
596 subjected to NOR test on Day 28 (A). NOR indices in Phases I (B), II (C), and III (D) were calculated.
597 Data are represented as means \pm SEM. Differences were considered significant at $^*p < 0.05$ and $^{**}p <$
598 0.01.

599 **Figure 3. Hypothalamic neuropeptide mRNA levels.** (A) mRNA levels of agouti-related peptide
600 (AgRP), neuropeptide Y (NPY), proopiomelanocortin (POMC), cocaine- and amphetamine-regulated
601 transcript (CART), corticotropin-releasing factor (CRF), urocortin (Ucn) 1, Ucn2, Ucn3, arginine
602 vasopressin (AVP), oxytocin (OXT), and orexin were measured in hypothalami isolated on Day 30.
603 (B) Correlation between mRNA expressions of AgRP or Ucn2 and Phase III NOR indices are
604 represented with respective Pearson correlation coefficients. Data are represented as means \pm SEM (n
605 = 8 – 10). Differences were considered significant at $*p < 0.05$ and $**p < 0.01$.

606 **Figure 4. Immunostaining for c-Fos and AgRP.** Coronal sections of arcuate nuclei were stained with
607 anti-mouse c-Fos and/or anti-AgRP antibody. (A) Representative images of c-Fos-positive cells (white
608 arrowheads) in arcuate nucleus (white dashed line) when subjected to immunofluorescent staining. (B)
609 c-Fos-positive cells were enumerated on one side of each of the six hypothalamus tissue sections per
610 mouse in Sham/solid ($n = 8$), Sham/powder ($n = 6$), and Malocclusion/powder group ($n = 7$). The
611 numbers of c-Fos-positive cells were normalized by DAPI-positive cells. Data are represented as means
612 \pm SEM. Differences were considered significant at $**p < 0.01$. (C) Representative images of AgRP-
613 and c-Fos-, DAPI- and c-Fos-, AgRP- and DAPI-, AgRP- and PGP9.5-, and DAPI- and PGP9.5-
614 positive cells in arcuate nuclei subjected to immunofluorescent staining (white arrows).

615 **Figure 5. NOR indices of mice subjected to intracerebroventricular (icv) anti-AgRP antibody**
616 **administration.** Anti-AgRP antibody (0.1 $\mu\text{g}/2 \mu\text{L}/\text{mouse}$) was administered intracerebroventricularly
617 to all mice from Day 25 to Day 30. NOR tests were performed 30 min after icv administration on Day
618 30. (A–C) Phase I (A), Phase II (B), and Phase III (C) NOR indices of Malocclusion/powder group
619 mice administered vehicle ($n = 5$) or antibody ($n = 5$). Data are represented as means \pm SEM.
620 Differences were considered significant at $**p < 0.01$.

Figure 1

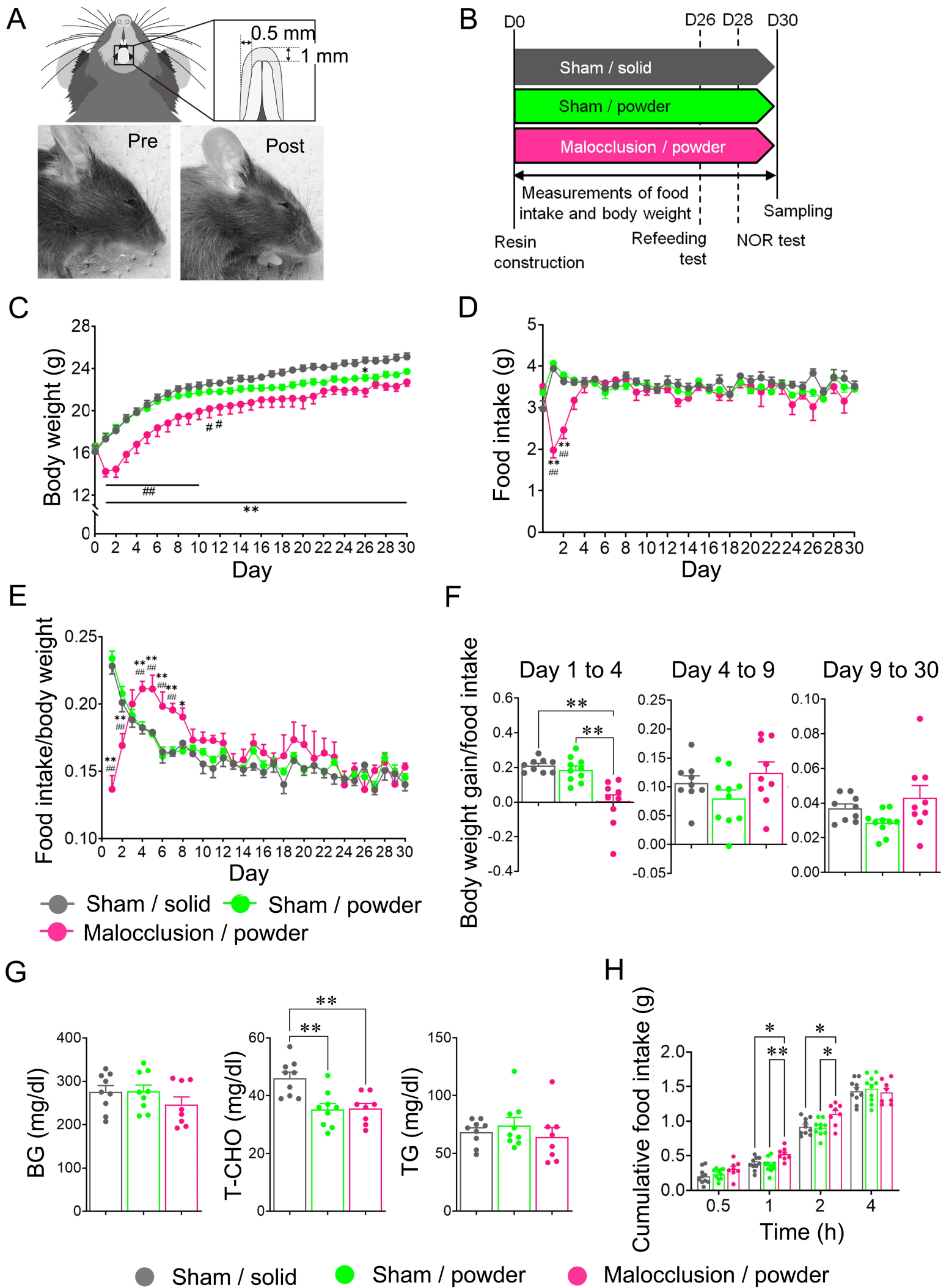


Figure 2

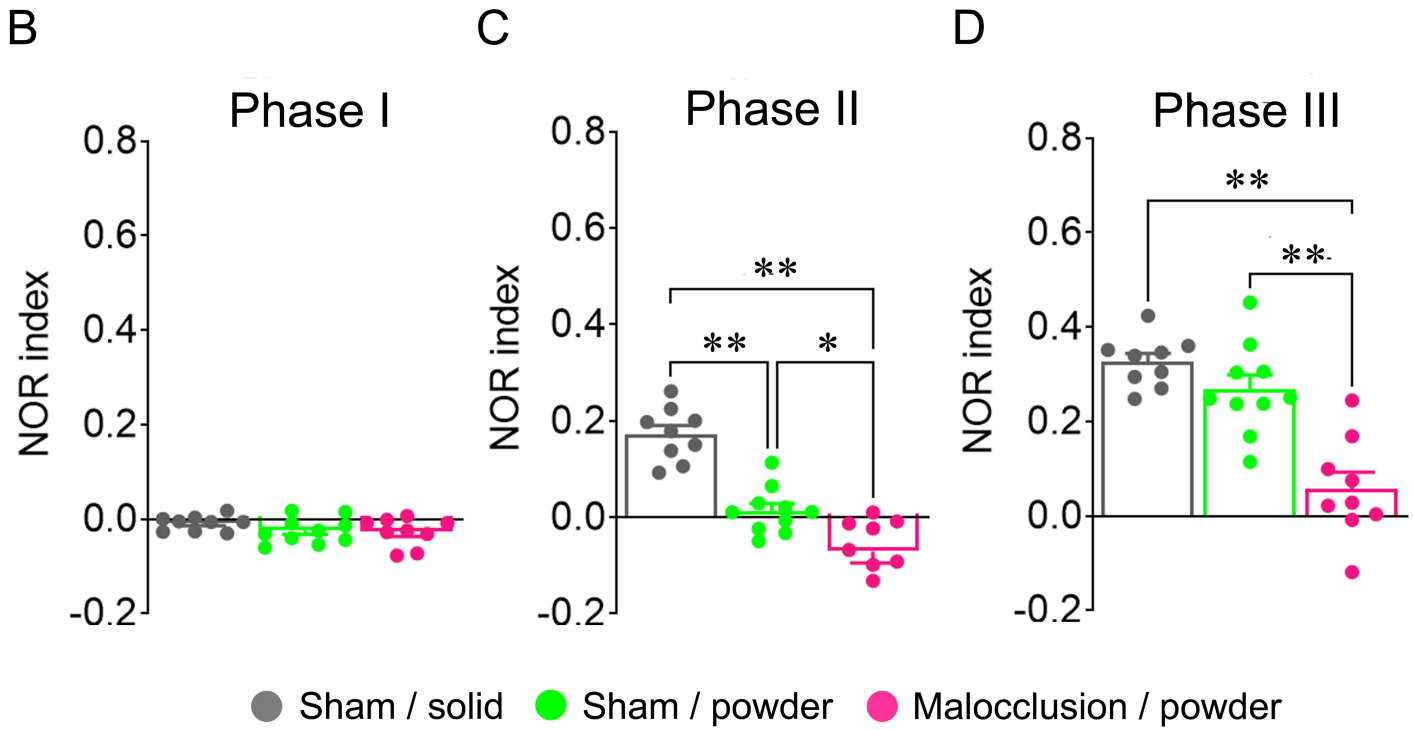
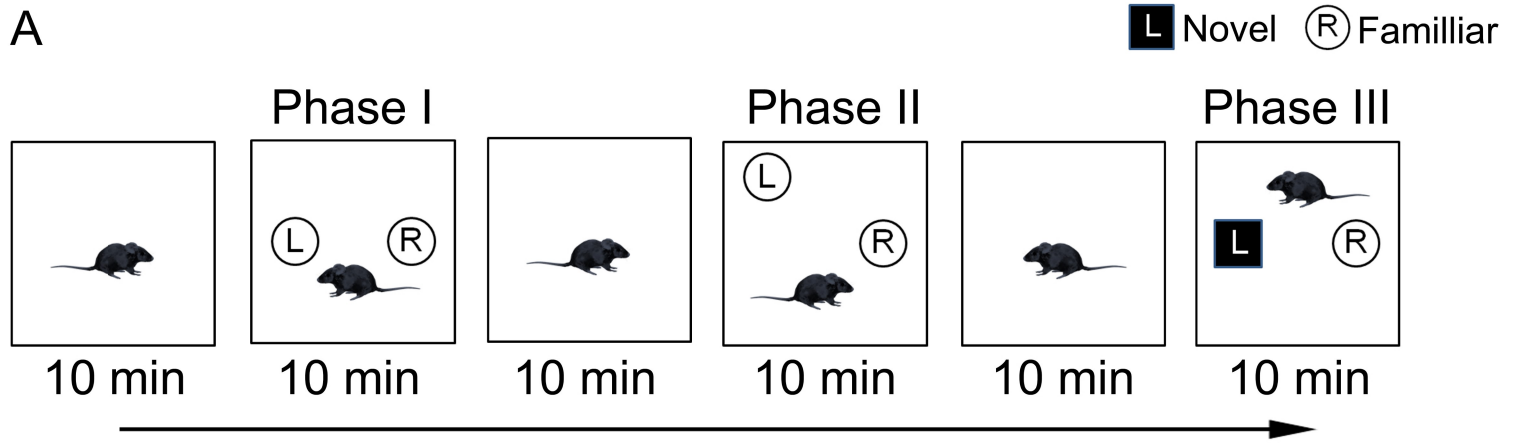
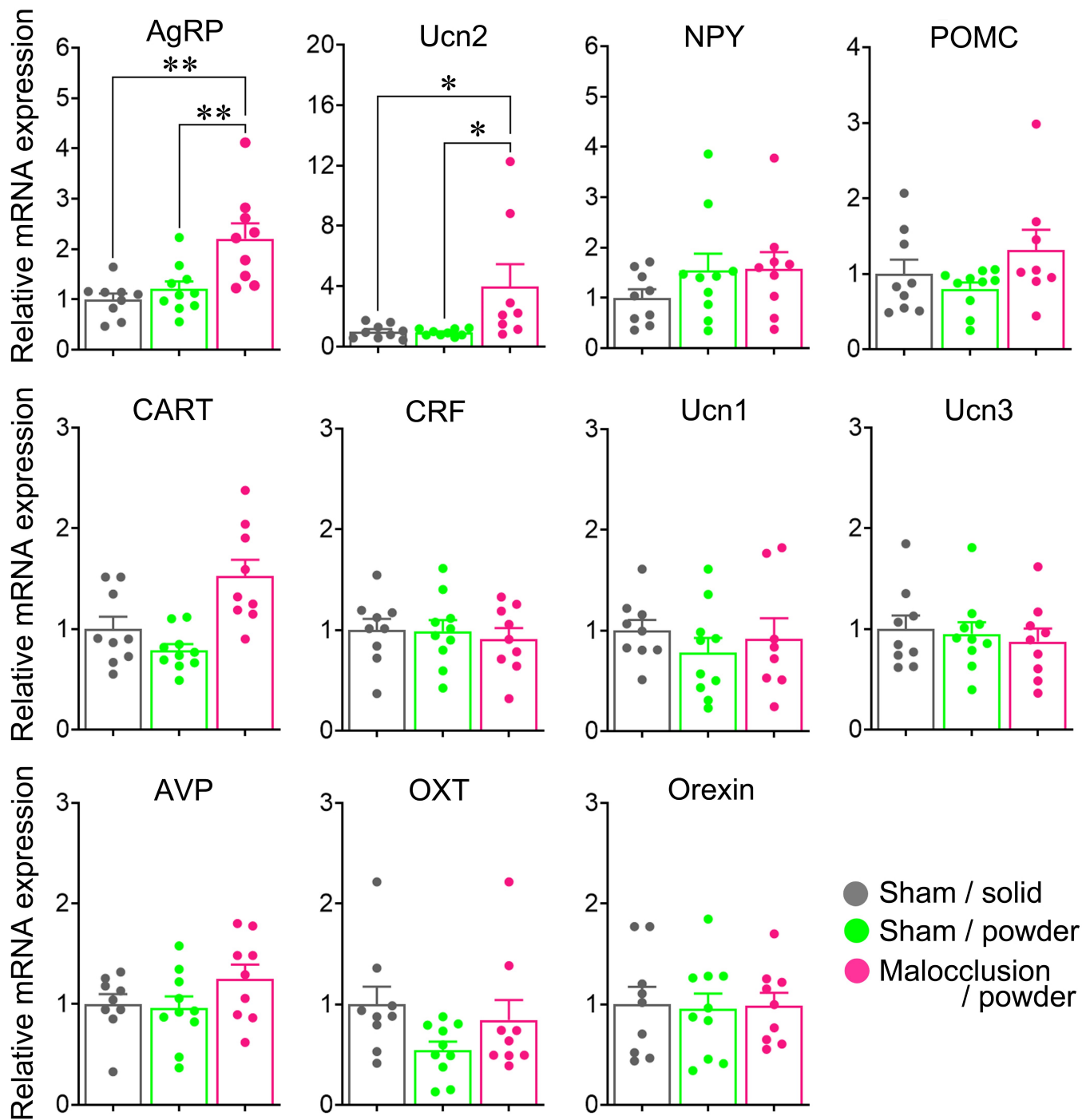


Figure 3

A



B

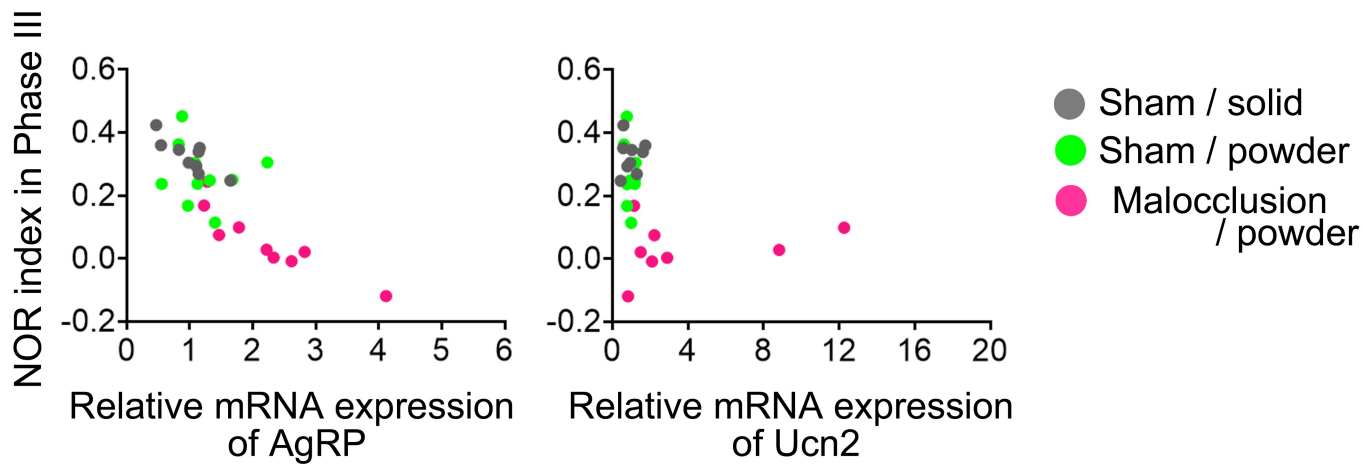


Figure 4

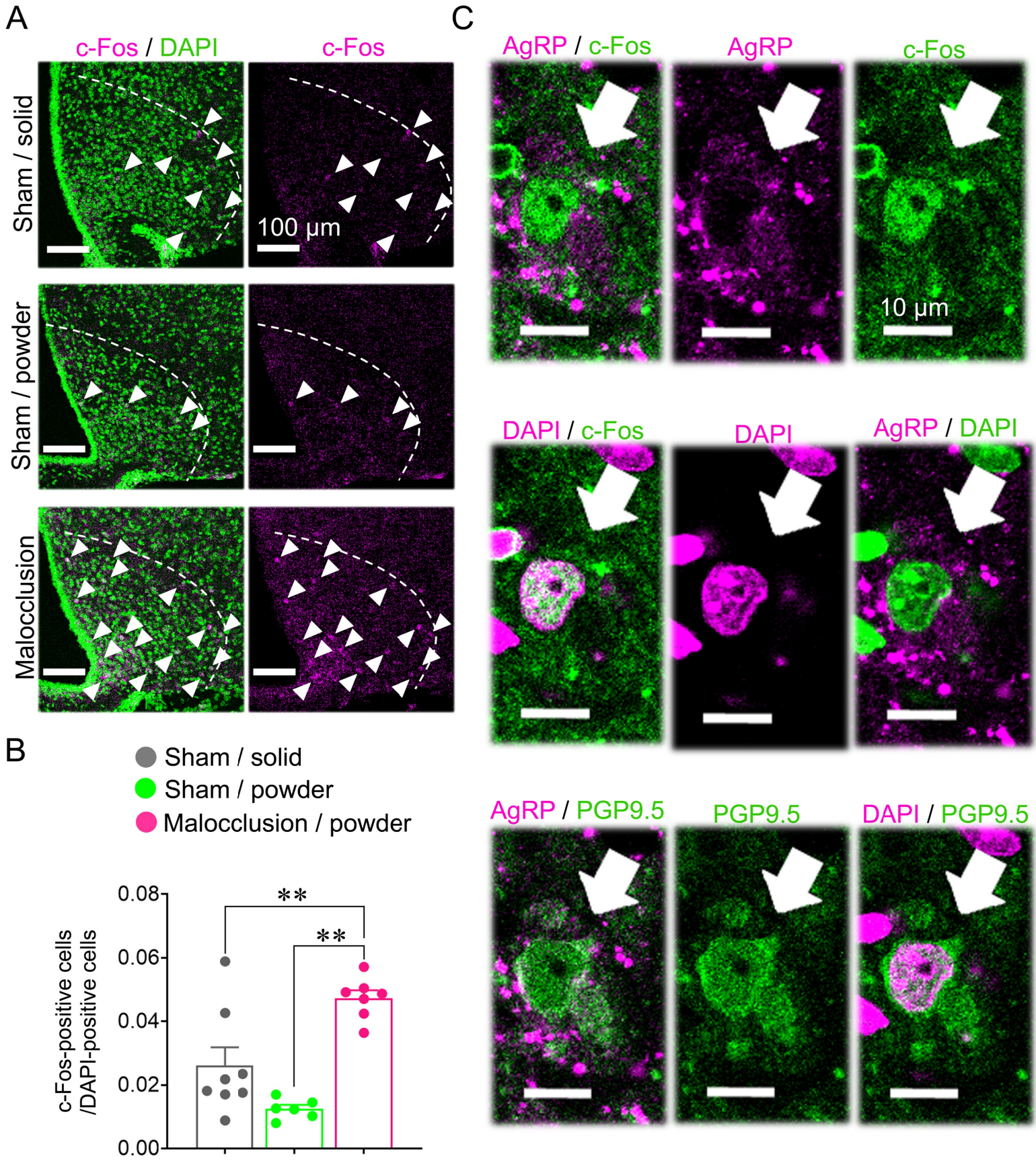


Figure 5

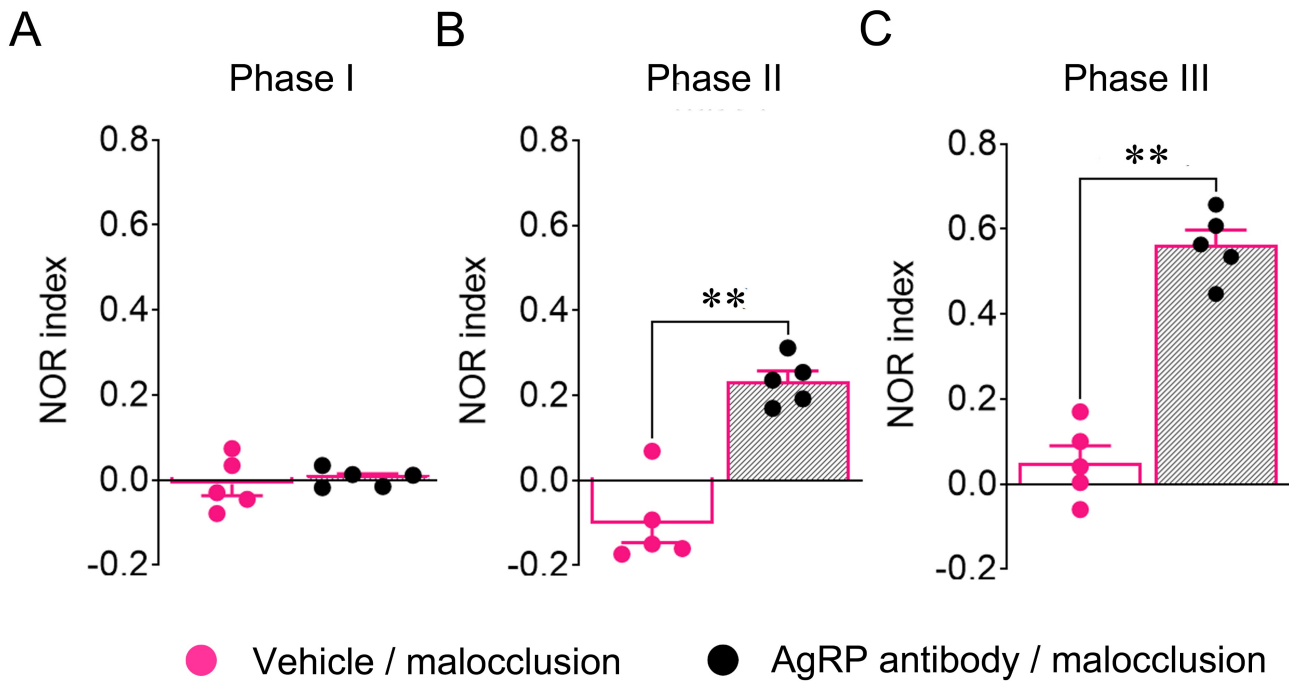
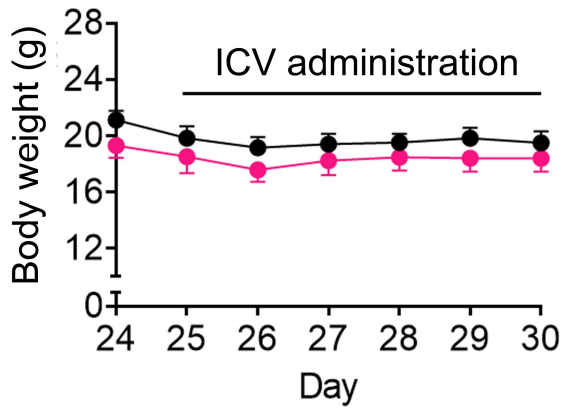


Figure S1

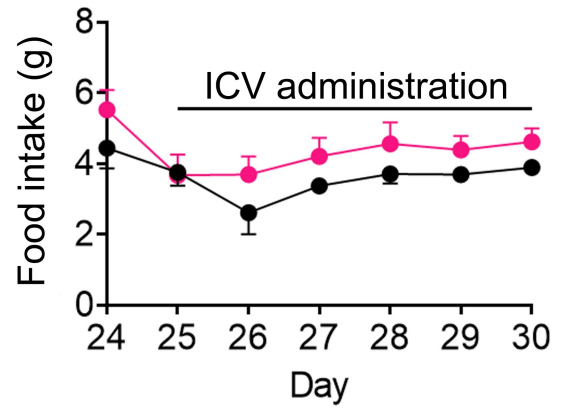
A

- Vehicle / malocclusion
- AgRP antibody / malocclusion



B

- Vehicle / malocclusion
- AgRP antibody / malocclusion



Supplementary Material

Malocclusion Impairs Cognitive Behavior via AgRP Signaling in Adolescent Mice

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4 **1 Supplementary Methods**

5 **1.1 Animals**

6 In the present study, n refers to the number of animals. Each mouse was considered to be an
7 experimental unit within this study. We set the humane endpoint as follows. If abnormal behavior,
8 such as immobility or tremor, was observed during the experiment, animals were euthanized with an
9 overdose of anesthesia. Abnormal mice were not observed in the present study. Although 92 mice
10 were used in this study, the resin of 3 mice and icv cannulae of 2 mice became dislodged during the
11 operation. Therefore, 87 mice were included in this study. The number of mice in each experiment
12 was as follows. We used the same mice in body weight and food intake (Fig. 1C-E), measurements of
13 glucose, total cholesterol, and triglyceride in perioheral blood, NOR behavior (Fig. 2B-D), and
14 hypothalamic mRNA level (Fig. 3) experiments. These experiments started with 29 mice (Sham/solid
15 group: 9, Sham/solid powder: 10, and Malocclusion/powder group: 10) but 1 mouse in the
16 Malocclusion/powder group was excluded from the analysis because the resin fell off in the middle
17 of the experiment. And more, 1 mouse in the Sham/powder and 1 mouse in the Malocclusion/powder
18 group were excluded from the blood analysis because sampling was fault. all The cumulative post-
19 fasting food intake experiment started with 30 mice (Sham/solid group: 10, Sham/powder group: 10,
20 and Malocclusion/powder group: 10) but 2 mice in the Malocclusion/powder group were excluded
21 from the analysis because their resins fell off in the middle of the experiment (Fig. 1F). We randomly
22 selected 21 mice (Sham/solid group: 8, Sham/powder group: 6, and Malocclusion/powder group: 7)
23 from mice used in the cumulative post-fasting food intake experiment for immunostaining (Fig. 4).
24 The icv anti-AgRP antibody administration experiment started with 12 mice, but 2 mice were
25 excluded from the analysis because their ICV cannulae fell off in the middle of the experiment.
26 Finally, each of the five mice treated with vehicle and AgRP antibody was examined using the NOR
27 test (Fig. 5).

28 **2 Supplementary Figures and Tables**

29 **2.1 Supplementary Tables**

30

Table S1. Primers used in RT-qPCR

		Forward	Reverse
neuropeptide Y (NPY)	NM_023456	CGCTCTGCGACACTACATCAAT	TGAGATGAGGGTGGAACCTGG
agouti-related peptide (AgRP)	NM_007427	GGACTGAGCATAAAGATGGCATGA	TGTAGCCAGGGCATGAGGTG
proopiomelanocortin (POMC)	NM_001278581	AATTACGTGGGTTATAGGACAGGAC	CCCTGAGCGACTGTAGCAGA
cocaine- and amphetamine-regulated transcript (CART)	NM_013732	GACATCTACTCTGCCGTGGATGA	TTCTTGCAACGCTTCGATCTG
corticotropin-releasing factor (CRF)	NM_205769	CAGAGCCCAAGTACGTTGAGAG	GCTCTCTTCTCCTCCCTTGTA
urocortin1 (Ucn1)	NM_021290	CATCTTGCACTGGGCAGACACT	AAGCTGTGCCAAGAGCAGCAAC
urocortin2 (Ucn2)	NM_145077	GACAGCCACAAAGCTGGACAGTA	GGCTCAGAAGCATGGCAAGA
urocortin3 (Ucn3)	NM_031250	CCACTCCAGAGCAAAGTCCACTTAC	GCTCAGCAAGGGCACATCTTC
arginine vasopressin (AVP)	NM_009732	TCTCTGACATGGAGCTGAGACAG	AGGGCAGGTAGTTCTCCTCCT
oxytocin (OXT)	NM_012996	TGCCAGGAGGAGAACTACCTG	TATCCCAGAAAGTGGGCTCAG
orexin	NM_010410	CGTAACTACCACCGCTTTAGCA	TGCCATTTACCAAGAGACTGACA
glyceraldehyde-3-phosphate dehydrogenase (GAPDH)	NM_008084	TGTGTCCGTCGTGGATCTGA	TTGCTGTTGAAGTCGCAGGAG

2.2 Supplementary Figures

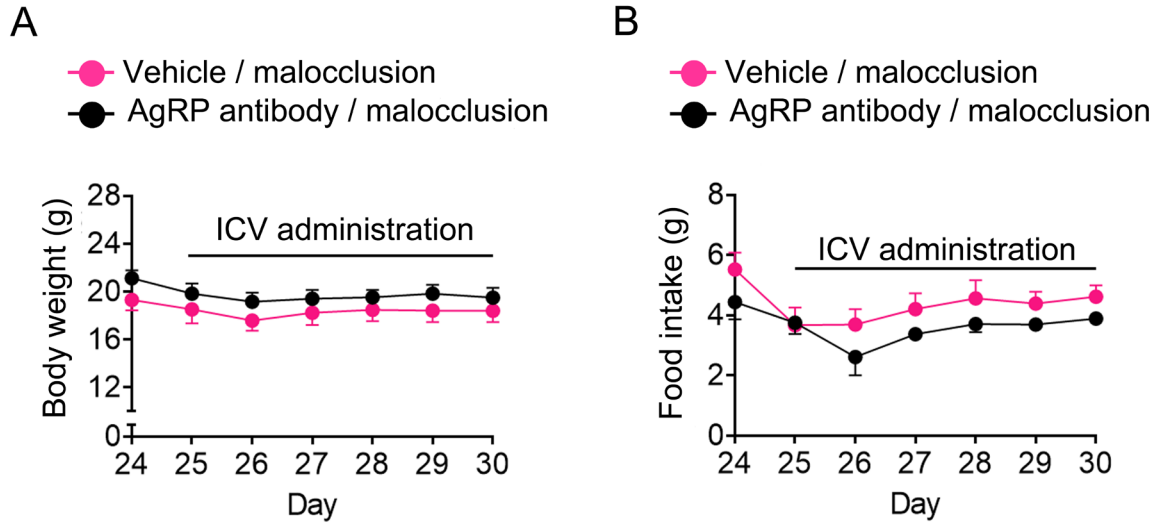


Figure S1. Time courses of body weight and food intake in mice repeatedly subjected to intracerebroventricular (icv) anti-AgRP antibody administration. (A, B) Body weight (A) and food intake (B) of mice in the Malocclusion/powder group after icv administration of vehicle ($n = 5$) or anti-AgRP antibody ($n = 5$). Data are shown as means \pm SEM. Bar indicates icv administration period.