Mechanisms underlying the modulation of the L-type Ca^{2+} channel by hydrogen peroxide in guinea-pig ventricular myocytes

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L. Yang carried our most of the experiments and wrote the manuscript.

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Abstract

Although Cav1.2 Ca^{2+} channels are modulated by reactive oxygen species (ROS), the underlying mechanisms are not fully understood. In this study, we investigated effects of hydrogen peroxide (H_2O_2) on the Ca^{2+} channel using a patch-clamp technique in guinea-pig ventricular myocytes. Externally applied H_2O_2 (1 mM) increased Ca^{2+} channel activity in the cell-attached mode. A specific inhibitor of Ca^{2+}/calmodulin-dependent protein kinase II, (CaMKII) KN-93 (10 μM), partially attenuated the H_2O_2-mediated facilitation of the channel, suggesting both CaMKII-dependent and independent pathways. However, in the inside-out mode, 1 mM H_2O_2 increased channel activity in a KN-93-resistant manner. Since H_2O_2-pretreated calmodulin did not reproduce the H_2O_2 effect, the H_2O_2 target was presumably assigned to the Ca^{2+} channel itself. A thiol-specific oxidizing agent mimicked and occluded the H_2O_2 effect. These results suggest that H_2O_2 facilitates the Ca^{2+} channel through oxidation of cysteine residue(s) in the channel as well as via the CaMKII-dependent pathway.

Keywords calcium channel, reactive oxygen species, H_2O_2, calmodulin, cardiac myocytes

Introduction

An alteration in the cell's redox state such as increased reactive oxygen species (ROS) production is associated with pathology [1-3]. In the heart, ROS as highly reactive compounds accumulate in tissues during myocardial ischemia/reperfusion and cause peroxidation of lipids and proteins [4], which play an important role in the pathogenesis of ischemia/reperfusion abnormalities, including myocardial stunning, irreversible injury, and reperfusion arrhythmias [5]. ROS-induced Ca^{2+} overload is one of the major causes of cardiomyocyte injury during ischemia/reperfusion [6]. Ca^{2+} overload induced by oxidation is thought to be mediated by increased Ca^{2+} release from the sarcoplasmic reticulum (SR) through the Ca^{2+} release channel (ryanodine receptor 2, RyR2) and decreased Ca^{2+} uptake by inhibiting Ca^{2+}-ATPase (SERCA) activity [7].

L-type (Cav1.2) Ca^{2+} channels (LTCCs) in the myocardium sarcolemma are the main route for Ca^{2+} influx into cells. Different from skeletal muscle, in cardiac myocytes, Ca^{2+} influx through LTCCs triggers Ca^{2+} release, determining the Ca^{2+} dynamics in the cardiac myocytes. Accumulating evidence shows that basal activity of LTCCs is modulated by cytoplasmic factors including protein kinase-mediated phosphorylation [8, 9], phosphatase-mediated dephosphorylation (8-9) and the interaction with Ca^{2+} and Mg^{2+} [10], lipids [11] and proteins [12, 13]. Recent studies suggest that the function of LTCC is crucially modulated by ROS during ischemia/reperfusion [14, 15]. Exposure of myocytes to a high concentration of H_2O_2 results in alteration of Ca^{2+} channel activity and cellular Ca^{2+} homeostasis [16]. However, the mechanism underlying the effects of oxidation is so far controversial since both inhibition and facilitation of LTCCs by oxidation have been suggested [17-20]. The mechanism of the ROS effect on LTCCs also remains elusive. For example, H_2O_2 has been suggested to facilitate LTCCs by activation of Ca^{2+}/CaM-dependent protein kinase II (CaMKII) through oxidation of methionine residues in CaMKII or increasing Ca^{2+} release through RyR (20). On the other hand, Tang *et al*. [19] have reported that H_2O_2-induced facilitation of LTCCs is mediated by glutathionylation of LTCCs.

To explore the effect of oxidation on cardiac LTCCs and the underlying mechanisms, the inside-out mode of patch-clamp technique is beneficial since the internal side of LTCC can be well controlled. We have previously found that LTCC activity is maintained with calmodulin (CaM) and ATP without run-down of the channel in the inside-out patches [12, 21-26]. In this study, using this method we have investigated the effect of H_2O_2 on the current through LTCCs. H_2O_2 was found to increase Ca^{2+} channel activity both in the cell-attached mode and the inside-out mode. The specific CaMKII inhibitor KN-93 partially attenuates the facilitation effect in the cell-attached mode, but had no effect in the inside-out mode. Our results suggest H_2O_2 facilitates LTCCs through CaMKII-dependent and independent pathways.

Materials and methods

Materials

MgATP, hydrogen peroxide (H_2O_2) tablet and 5,5'-dithiobis (2-nitrobenzoic acid) (DNTB) (cysteine residue oxidation reagent) were purchased from Sigma-Aldrich (St. Louis, MO, USA), KN-93 (CaMKII inhibitor) and KN-92 (inactive analog of KN-93) were from Calbiochem (San Diego, CA, USA) and Bay K 8644 (Ca^{2+} channel agonist) was from Wako (Osaka, Japan).

Preparation of single cardiac myocytes

Single ventricular myocytes were isolated from adult guinea-pig hearts by collagenase dissociation as described previously [27]. In brief, a female guinea pig (weight 300-500 g) was anesthetized with pentobarbital sodium (30 mg/kg, i.p.), and the aorta was cannulated in situ under artificial respiration. The dissected heart was mounted on a Langendorff apparatus and perfused with Tyrode solution for 3 min, then with nominally Ca^{2+}-free Tyrode solution for 5 min, and finally with Ca^{2+}-free Tyrode solution containing collagenase (0.08 mg/ml; Yakult) for 7–15 min. The collagenase was washed away with a high K^+, low Ca^{2+} solution (storage solution). The single ventricular myocytes were dispersed and filtered through a stainless steel

mesh (105 μm). 0.05 mg/ml protease (Type XIV, Sigma) and 0.02 mg/ml DNase I (Type IV, Sigma) were incubated with the myocytes to improve the success rate in attaining a gigaohm seal. The enzyme-treated myocytes were then washed twice by centrifugation (800 rpm for 3 min) and stored at 4^{o}C.

The experiments were carried out under the approval of the Committee of Animal Experimental, Kagoshima University.

Solutions

Tyrode solution contained (in mM) 135 NaCl, 5.4 KCl, 0.33 NaH_2PO_4, 1.0 $MgCl_2$, 5.5 glucose, 1.8 $CaCl_2$, and 10 HEPES-NaOH buffer (pH 7.4). The storage solution was composed of (in mM) 70 KOH, 50 glutamic acid, 40 KCl, 20 KH_2PO_4, 20 taurine, 3 $MgCl_2$, 10 glucose, 10 HEPES, and 0.5 EGTA and the pH was adjusted to 7.4 with KOH. The pipette solution contained (in mM) 50 $BaCl_2$, 70 tetraethylammonium chloride, 0.5 EGTA, 0.003 BAY K 8644, and 10 HEPES-CsOH buffer (pH 7.4). The basic internal solution consisted of (in mM) 90 potassium aspartate, 30 KCl, 10 KH_2PO_4, 1 EGTA, 0.5 $MgCl_2$, 0.5 $CaCl_2$, and 10 HEPES-KOH buffer (pH 7.4; free Ca^{2+} 80 nM, pCa 7.1). CaM and MgATP were dissolved in basic internal solution in the inside-out patch mode.

Preparation of CaM

The cDNA of human CaM cloned into the pGEX6P-3 vector (GE Healthcare Bioscience, Uppsala, Sweden) was expressed as a glutathione-S-transferase (GST) fusion protein in *Escherichia coli* BL21 and purified using glutathione–Sepharose 4B (GE Healthcare). The GST region was cleaved by PreScission Protease (GE Healthcare). The purity of CaM was confirmed by SDS-PAGE and quantified by the Bradford method (Thermo Fisher Scientific, Rockford, IL, USA) with bovine serum albumin as the standard and a correction factor of 1.69.

Patch clamp and data analysis

Barium current through LTCCs was recorded in the cell-attached and inside-out mode using the patch-clamp technique. For the recording in the cell-attached mode, the myocytes were perfused with the basic internal solution at 31–35°C using a patch pipette (2–4 MΩ) containing 50 mM Ba^{2+} and 3 μM Bay K8644. Bay K8644, a Ca^{2+} channel agonist, was used to prolong the open time of the channel to facilitate the experiments. Barium currents through LTCCs were elicited by depolarizing pulses from -70 to 0 mV for a 200 ms duration at a rate of 0.5 Hz. The currents were recorded with a patch-clamp amplifier (EPC-7; List, Darmstadt, Germany) and fed to a computer at a sampling rate of 3.3 kHz. The mean current (I) was measured and divided by the unitary current amplitude (i) to yield NPo, based on the equation $I = N \times Po \times i$, where N is the number of channels in the patch and Po is the time-averaged open-state probability of the channels. Data are presented as the mean ± S.E. Student's t-test or ANOVA with post-hoc Tukey HSD test was used to estimate statistical significance and values of $P < 0.05$ were considered as significant.

RESULTS

H_2O_2 facilitated Ca^{2+} channel in the cell-attached mode via CaMKII-dependent and independent pathways

We first examined the effect of H_2O_2 on the current through LTCCs in the cell-attached mode in guinea-pig ventricular myocytes. After recording the current for 2 minutes as a control, 1 mM H_2O_2 was applied in the perfusion solution. As shown in Fig. 1A and B, Ca^{2+} channel activity was rapidly increased without a change in the unitary current amplitude. Taking an average of six patches, the Ca^{2+} channel activity was increased to 206 ± 32 % of the control (Fig. 1D). This result confirmed the facilitating effect of H_2O_2 on LTCCs.

Facilitation of LTCCs by glutathionylation [19] or phosphorylation mediated by activated CaMKII [20] during oxidative stress has been proposed. To evaluate the possible CaMKII-dependent effect of H_2O_2, we incubated the cardiomyocytes with 10 μM KN-93, a specific CaMKII inhibitor, for 10 minutes before recording the current. Under this condition, it has

been reported that activity of CaMKII is nearly completely inhibited by KN-93 [28, 29]. As shown in Fig. 1C, KN-93 significantly attenuated H_2O_2-mediated facilitation (132 ± 15 % (n=9) vs 206 ± 32 % with no drug, P<0.05), while KN-92, an inactive analog of KN-93, partially attenuated the facilitation but it was not statistically significant (156 ± 13 % (n=5) vs 206 ± 32 % with no drug, P=0.30). These results suggested that not only CaMKII-dependent but also independent pathways were involved in H_2O_2-mediated facilitation of LTCCs.

H_2O_2 facilitated Ca^{2+} channel in the inside-out mode independently of CaMKII

To explore the mechanism of CaMKII-independent facilitation of LTCCs produced by H_2O_2, we investigated the H_2O_2 effect on LTCCs in the inside-out patches in which the Ca^{2+} channel activity was maintained by application of 1 μM CaM together with 3 mM ATP [12, 21-26]. In the inside-out patch mode, LTCCs were disconnected with cytoplasmic factors and perfused with an artificial solution with known composition. This was quite beneficial to examine direct effects of external reagents on LTCC. After the patch was excised and moved to a small inlet in the perfusion chamber which was connected to a microinjection system, CaM/ATP was immediately applied to induce Ca^{2+} channel activity, the single channel current was recorded for 3 minutes as a control current, then 1 mM H_2O_2 was added into the CaM/ATP solution. As shown in Fig. 2A, H_2O_2 significantly increased the CaM-induced Ca^{2+} channel activity in the inside-out mode. This facilitatory effect of H_2O_2 was concentration dependent up to 1-2 mM, and higher concentrations of H_2O_2 conversely inhibited Ca^{2+} channel activity presumably because of a non-specific deteriorating effect of H_2O_2 (Fig. 2B). These results suggested that H_2O_2 (<2 mM) facilitated Ca^{2+} channel activity in the inside-out patches via a direct modification of LTCC and/or its closely-located proteins such as CaM and CaMKII. To assess the possibility that CaMKII was still located near the channel and modulated channel activity in the excised patches, we examined the effect of KN-93 in the inside-out patches. As shown in Fig. 2C, KN-93 had only a small effect on the H_2O_2-mediated facilitation of LTCCs in the inside-out mode. In summary, channel activity was modulated by H_2O_2 from 144 ± 32 %

(control) to 272 ± 70 % in the absence of KN-93 (n=5), and from 139 ± 21 % to 231 ± 29 % in the presence of KN-93 (n=6). Although KN-93 seemed to slightly attenuate the increasing effect of H_2O_2 on channel activity, this difference was statistically insignificant. Thus KN93 did not significantly affect the H_2O_2-mediated facilitation of the Ca^{2+} channel in the inside-out mode.

H_2O_2 may also able to oxidize CaM and modulate the effect of CaM on Ca^{2+} channel activity. To assess this possibility, we examined the effect of oxidized CaM pretreated with 1 mM H_2O_2 at room temperature for 30 minutes. As shown in Fig. 3A and B, after Ca^{2+} channel activity was maintained by intact (untreated with H_2O_2) CaM + ATP in the inside-out patches, we substituted the oxidized CaM for the untreated CaM. Channel activity did not change significantly, suggesting that CaM was not oxidized or oxidized CaM, if any, had similar potency as the untreated CaM on activity of LTCC (Fig. 3C). This result suggested that oxidation of CaM was not involved in the H_2O_2-mediated facilitation of LTCCs, and thus a direct oxidation of LTCCs might be a possible mechanism for the facilitation in the inside-out mode.

Cysteine residues in the Ca^{2+} channel are involved in H_2O_2-mediated facilitation

The α1C subunit of LTCC contains 38 cysteine residues and 36 methionine residues in the cytoplasmic chains, which are potentially subject to oxidation modification. To identify the amino acid residue which was oxidized by H_2O_2 and responsible for the H_2O_2-mediated facilitation of LTCCs, a specific oxidizing agent of cysteine residues 5,5'-dithiobis-(2-nitrobenzoic acid) (DTNB) was applied in place of H_2O_2. As shown in Fig. 4, 1 mM DNTB significantly increased Ca^{2+} channel activity maintained by CaM (from 129 ± 22 % up to 184 ± 21 %, n=5), suggesting that oxidation of cysteine residue(s) was responsible, at least partially, for the H_2O_2-mediated facilitation of LTCCs. Since there is no specific oxidizing agent of methionine residues available, we examined the effects of H_2O_2 on LTCC after application of DTNB. Application of H_2O_2 + CaM after DTNB + CaM only slightly increased

Ca^{2+} channel activity and was statistically insignificant (Fig. 5). These results suggested that oxidation of cysteine residue(s) was the major cause of the H_2O_2-mediated facilitation of LTCCs in the inside-out patches.

Discussion

In the present study, we investigated the effect of hydrogen peroxide (H_2O_2) on the L-type Ca^{2+} channel (LTCC) of cardiac myocytes in the cell-attached and the inside-out mode. We found that H_2O_2 facilitates cardiac LTCCs through two pathways: 1) direct oxidation of cysteine residue(s) of the channel; 2) indirect pathways via activation of CaMKII.

Changes of the redox state in the cardiac myocytes play an important role in heart diseases. LTCC as a major regulator of cardiac function is subject to redox modification. Although there is accumulating evidence supporting that reactive oxygen species (ROS) modulate the function of LTCCs, the results of these studies are controversial: Oxidizing agents are reported to inhibit the human and rabbit cardiac LTCC expressed in HEK293 cells [17, 18]. In isolated guinea-pig ventricular myocytes, oxidation decreased the current through LTCC [30]. On the other hand, a decrease in cellular superoxide and H_2O_2 is associated with a decrease in the current of native [31, 32] and expressed LTCCs [17, 33], while the thiol-specific oxidizing agent DNTB increases the LTCC current [34]. The complicated interactions between LTCC and uncontrolled cytoplasmic factors may be partially responsible for the uncertainty surrounding the effects of oxidation on LTCCs. The present study took advantage of the inside-out mode of patch-clamp technique in which most of the cytoplasmic factors were washed out [12, 21-26]. Our results show that H_2O_2 facilitates LTCC at concentrations up to 2 mM and inhibits the channel at higher concentrations. This finding may partly account for the diverse results in the previous studies.

The underlying mechanisms of modulation of the Ca^{2+} channel by H_2O_2 are not completely clear. Song *et al*. [20] have reported that H_2O_2-mediated facilitation of the Ca^{2+} channel through activation of CaMKII can be activated either by Ca^{2+}/CaM or oxidation of methionine

residues in CaMKII. Thus, H_2O_2 is involved in both activation processes: 1) H_2O_2 enhances Ca^{2+} release from SR by increasing RYR activity; 2) Oxidation of methionine residues in CaMKII protein (amino acid numbers: 281 and 282 in mouse) sustains the kinase activity independent of Ca^{2+}/CaM [3]. However, Tang *et al*. [19] suggest that H_2O_2 facilitates the Ca^{2+} channel through direct glutathionylation of the channel protein. It is difficult to distinguish the direct effect of H_2O_2 from an indirect one when examining whole cells. In the present study, we employed a method to record LTCC current in the inside-out mode in which channel activity was maintained by CaM/ATP in the internal solution [12, 21-26]. Our results show that the CaMKII specific inhibitor KN-93 did not completely attenuate H_2O_2-mediated facilitation of LTCC in the cell-attached mode, suggesting that H_2O_2-mediated facilitation is mediated by not only a CaMKII-dependent pathway but also CaMKII-independent pathways. The finding that KN-93 does not inhibit the H_2O_2 effect in the inside-out mode indicated that the CaMKII-mediated facilitation is absent in the inside-out mode, and hence implies that CaMKII might not be attached to LTCC or, if present, it might not be in a state sensitive to oxidation in our inside-out patches.

Thus our results indicate that, in addition to the CaMKII-dependent pathway, there is an additional CaMKII-independent pathway for the H_2O_2-mediated facilitation of LTCC. Since most intracellular proteins are washed out in the inside-out patches, it is likely that direct oxidation of LTCC or its associated proteins by H_2O_2 might be involved in the facilitation of LTCC. Since H_2O_2-pretreated CaM does not mimic the facilitatory effect of H_2O_2, oxidation of CaM does not account for the mechanism of facilitation. This is consistent with the fact that human CaM does not contain any cysteine residues. Thus, it seems most likely that the Ca^{2+} channel protein itself undergoes direct oxidation by H_2O_2 as the CaMII-independent pathway of the LTCC facilitation.

Both cysteine and methionine residues are subject to oxidation by H_2O_2. The pore-forming subunit α1C of cardiac LTCC is rich in cysteine and methionine residues in the cytoplasmic chains [35]. Our findings that the specific cysteine oxidizing agent DTNB mimics

the H_2O_2 effect and that the effect of subsequently applied H_2O_2 is largely occluded suggests that cysteine residue(s) are involved in the H_2O_2-mediated facilitation. However, this does not exclude a possible involvement of a methionine residue(s). Future work should determine the oxidation sites responsible for the H_2O_2-mediated modulation of LTCC. In conclusion, LTCC may undergo ROS-mediated modification via the direct oxidation of LTCC as well as the indirect pathways involving CaMKII activation. This would be relevant for the understanding of ROS-mediated regulation of ion channels and Ca^{2+} overload and arrhythmogenesis during oxidation stress on the heart.

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Conflict of interest

The authors declare that they have no conflict of interest.

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Figure legends

Fig. 1 H_2O_2 facilitates L-type Ca^{2+} channel activity in cell-attached mode. **A** Time course of channel activity (*NPo*) recorded in cell-attached mode before and after application of 1 mM H_2O_2. **B** Examples of current traces of the Ca^{2+} channels before (a) and after (b) application of H_2O_2 taken at the times indicated in A. **C** Effect of 1 mM H_2O_2 on channel activity (*NPo*) recorded in the cell-attached mode in the presence of 10 μM KN-93. **D** Summary of the normalized activity of the Ca^{2+} channel treated with H_2O_2 with no drug (n=6), and with 10 μM KN-93 (a specific CaMKII inhibitor) (n=9) or KN-92 (an inactive form of KN-93) (n=5). Mean channel activity (60 traces) in each patch was normalized to the corresponding control value, averaged in the same group, and shown as mean ± S.E. * $P<0.05$ and ** $P<0.01$ vs. control (Student's t-test), and # $P<0.05$ and NS not significant vs. H_2O_2 without drug (ANOVA and Tukey HSD test).

Fig. 2 H_2O_2-mediated facilitation of Ca^{2+} channel activity in inside-out mode. **A** and **C** Time course of channel activity (*NPo*) recorded in cell-attached (c.a.) mode followed by inside-out (i.o.) mode, in which channel activity was maintained with 1 μM CaM + 3 mM ATP, and then 1 mM H_2O_2 without (A) or with (C) 10 μM KN-93 was additionally applied as indicated by the boxes in each graph. ATP was included throughout the experiments in the inside-out mode. **B** Concentration-dependent effect of H_2O_2. Normalized channel activity was plotted against concentration of H_2O_2. Data (n=4-6) were fitted with a combined Hill's equation as:

$$(100 + A\cdot[H_2O_2]\text{^}n_f/(Kd_f\text{^}n_f + [H_2O_2]\text{^}n_f))\cdot(Kd_i\text{^}n_i/(Kd_i\text{^}n_i + [H_2O_2]\text{^}n_i),$$

where $[H_2O_2]$ is the concentration of H_2O_2, A the extent of facilitation, Kd_f and Kd_i apparent dissociation constants, and n_f and n_i Hill's numbers for facilitation and inhibition, respectively. The fitted curve was drawn with A=147, Kd_f =0.68 mM, n_f =2, Kd_i =3.11 mM and n_i =3 (r^2=0.983). **D** Summary of the normalized channel activity in the presence of CaM + ATP in

i.o. mode before and after addition of H_2O_2 ± KN-93 (n=5-6). Data are shown as mean ± S.E. * P<0.05 vs. CaM (t-test), NS not significant (ANOVA).

Fig. 3 Facilitation of Ca^{2+} channel by H_2O_2 is not because of oxidation of CaM. CaM was pre-incubated with 1 mM H_2O_2 for 30 min and then applied to the Ca^{2+} channels. **A** Time course of channel activity recorded first in cell-attached (c.a.) mode followed by inside-out (i.o.) mode with 1 μM non-treated (intact) CaM + 3 mM ATP, followed by substitution with H_2O_2-treated CaM (1 μM). **B** Example of current traces for the control in c.a. mode (a), with CaM + ATP in i.o mode (b) and H_2O_2-treated CaM + ATP (c), at the time period indicated in A. **C** Summary of normalized channel activity induced by CaM + ATP (n=7) and H_2O_2-treated CaM + ATP (n=6). Data are shown as mean ± S.E. *** P<0.001 vs. control (c.a.), NS not significant (ANOVA and Tukey HSD test).

Fig. 4 DTNB, a specific oxidant of cysteine residues, facilitates Ca^{2+} channel activity in inside-out mode. A Time course of channel activity recorded first in cell-attached (c.a.) mode followed by inside-out (i.o.) mode. After patch excision, 1 μM CaM + 3 mM ATP was applied to maintain channel activity, and then DTNB (1 mM) was additionally applied. **B** Example of current traces for the control in c.a. mode (a), with CaM + ATP in i.o mode (b) and with CaM + ATP + 1 mM DTNB (c) at the time period indicated in A. **C** Summary of normalized channel activity in the presence of CaM + ATP and CaM + ATP + DTNB (n=5). Data are shown as mean ± S.E. * P<0.05 vs. control (t-test).

Fig. 5 A Time course of channel activity recorded first in cell-attached (c.a.) mode followed by inside-out (i.o.) mode. After patch excision, 1 μM CaM + 3 mM ATP was applied to maintain channel activity, then DTNB (1 mM) was added, and finally H_2O_2 (1 mM) was additionally applied. **B** Summary of channel activity induced by CaM + ATP + DTNB (n=5) and CaM + ATP + H_2O_2 (n=6), normalized to the activity values obtained for conditions of

CaM + ATP. Data are shown as mean ± S.E. NS not significant (t-test).

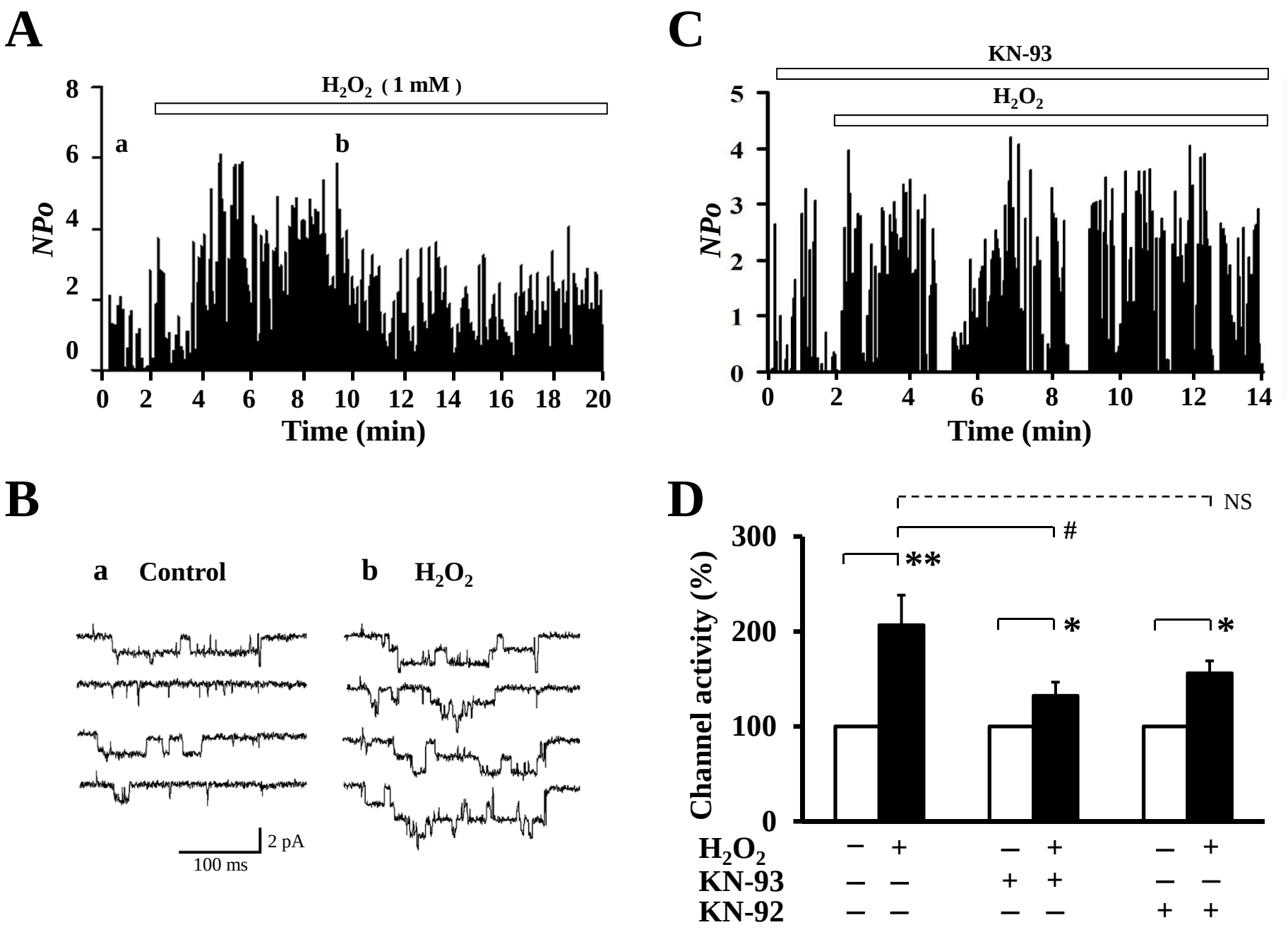
A
H_2O_2 (1 mM)
a
b
NPo
Time (min)
B
a Control
b H_2O_2
2 pA
100 ms
C
KN-93
H_2O_2
NPo
Time (min)
D
NS
#
**
*
*
Channel activity (%)
H_2O_2
KN-93
KN-92

Fig.1

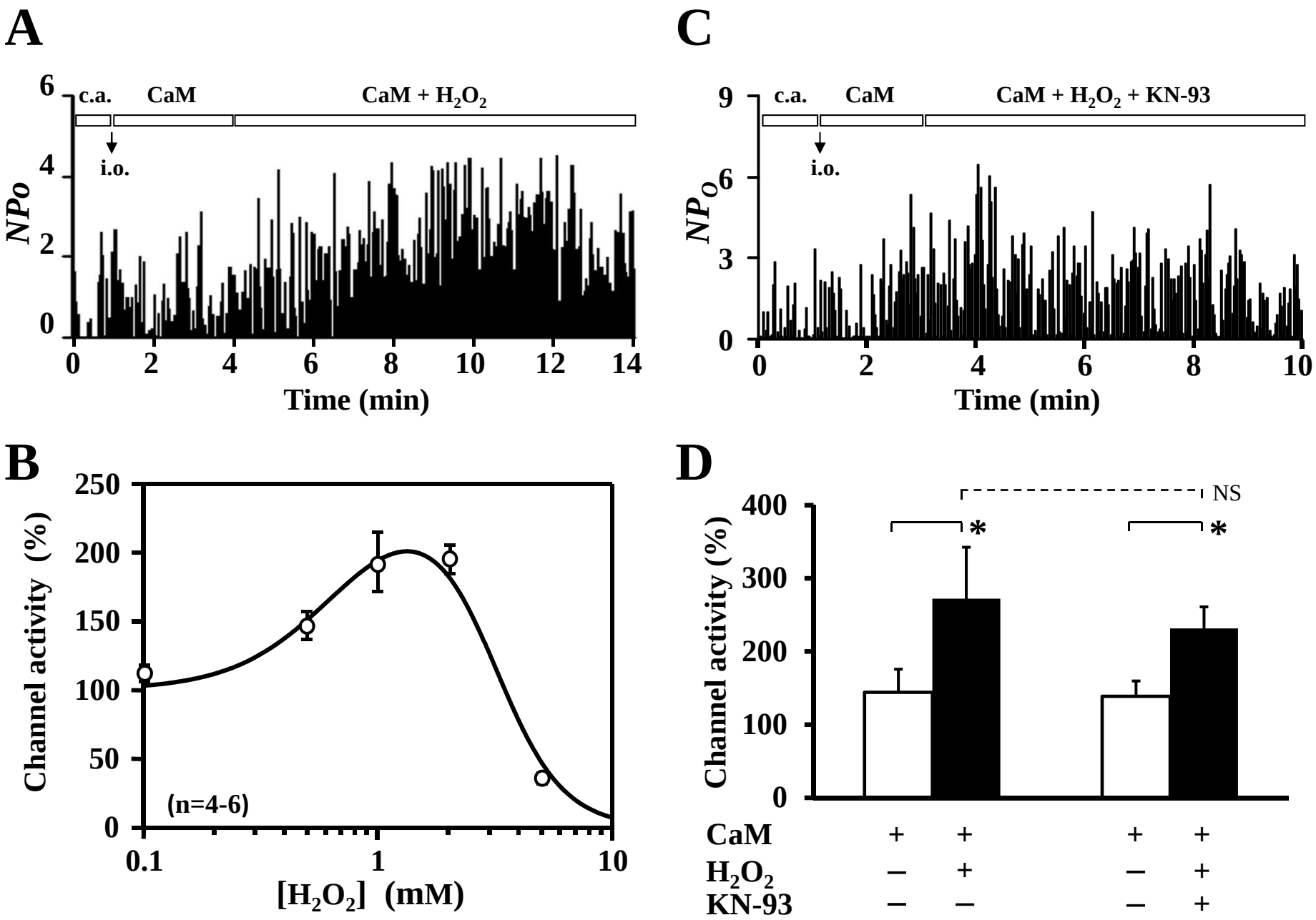
A
c.a.
CaM
CaM + H2O2
i.o.
NPo
Time (min)
C
c.a.
CaM
CaM + H2O2 + KN-93
i.o.
NPO
Time (min)
B
Channel activity (%)
(n=4-6)
[H2O2] (mM)
D
Channel activity (%)
NS
CaM
H2O2
KN-93

Fig.2

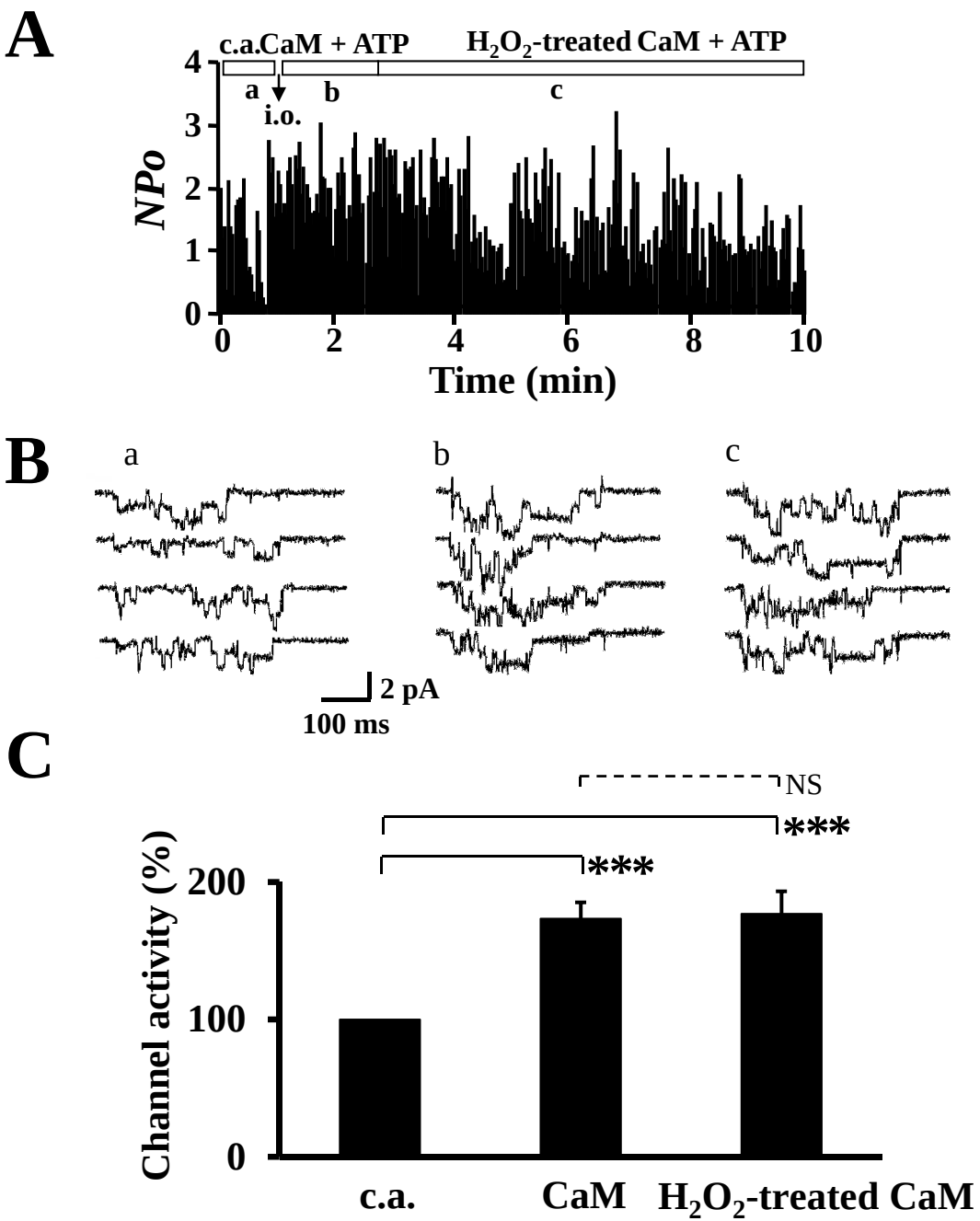
A
c.a.CaM + ATP
H2O2-treated CaM + ATP
a
b
c
i.o.
NPo
Time (min)
B
a
b
c
2 pA
100 ms
C
NS
***
***
Channel activity (%)
c.a.
CaM
H2O2-treated CaM

Fig.3

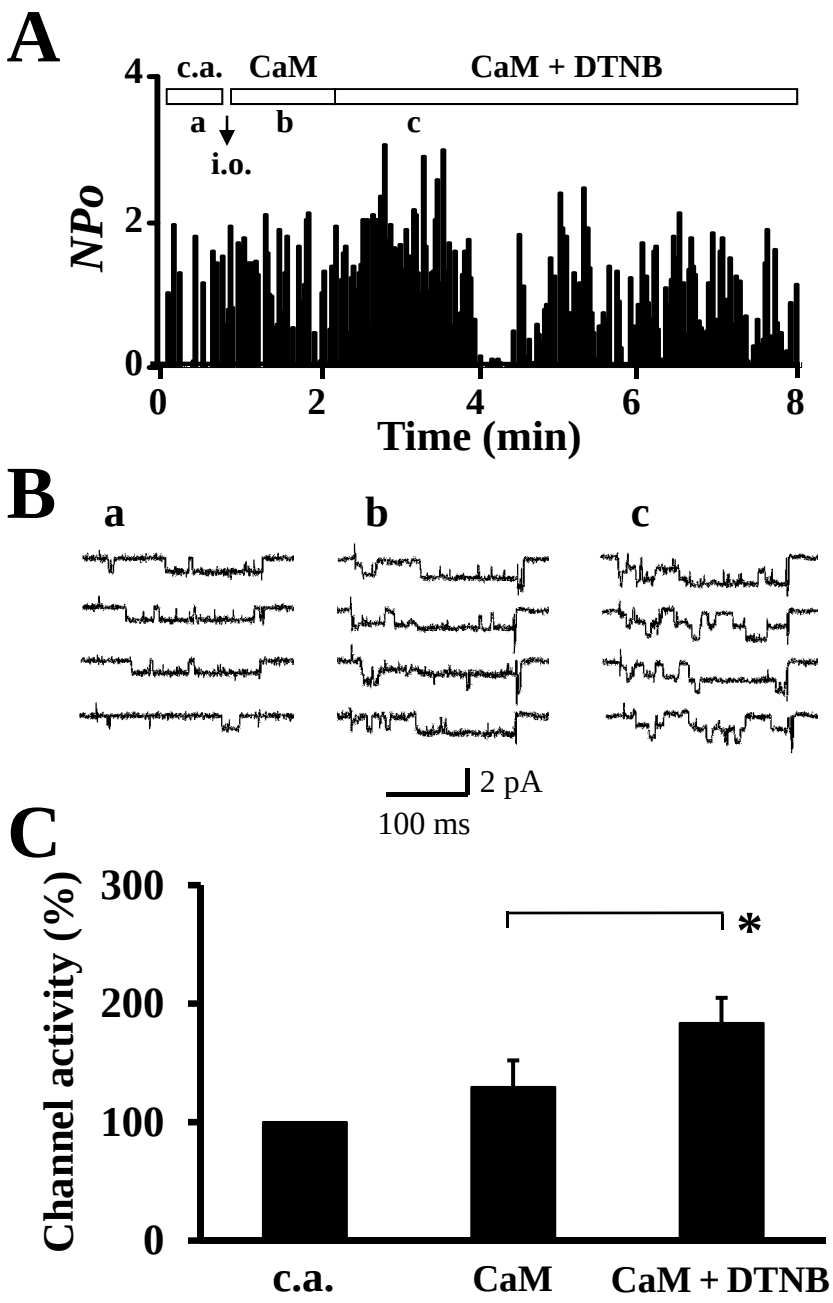
A
c.a.
CaM
CaM + DTNB
a
b
c
i.o.
NPo
4
2
0
0
2
4
6
8
Time (min)
B
a
b
c
2 pA
100 ms
C
Channel activity (%)
300
200
100
0
*
c.a.
CaM
CaM + DTNB

Fig.4

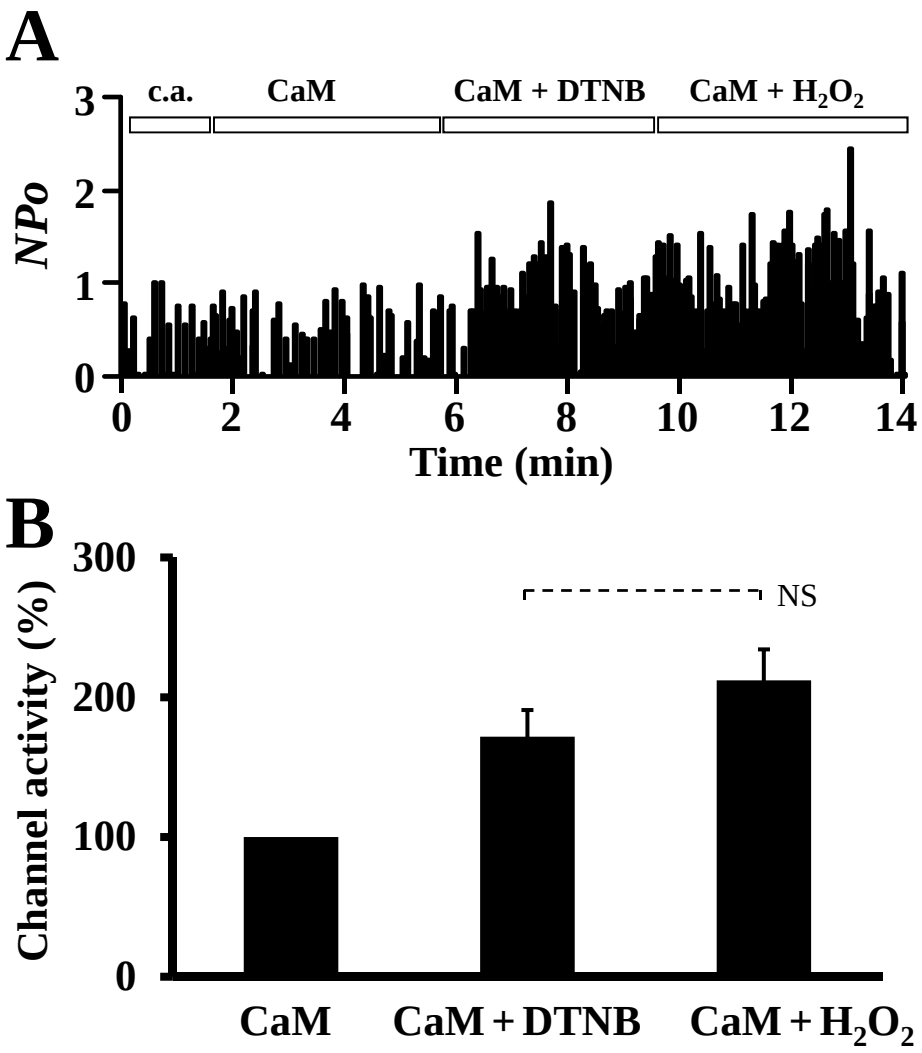
A
c.a.
CaM
CaM + DTNB
CaM + H2O2
NPo
3
2
1
0
0
2
4
6
8
10
12
14
Time (min)
B
Channel activity (%)
300
200
100
0
NS
CaM
CaM + DTNB
CaM + H2O2

Fig.5